

Past, present & future of mycoplasmas in chickens & turkeys

A. Feberwee

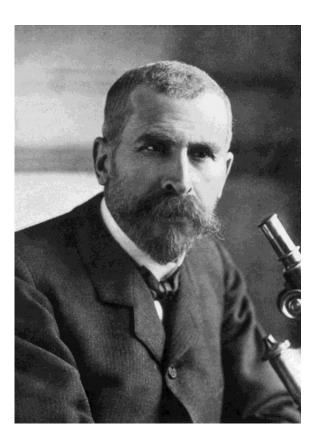
DVM, PhD, EBVS© European Specialist in Poultry Veterinary Science Specialist Poultry Health GD Animal Health, Deventer, the Netherlands



Introduction







The microbe (virus?) of pleuropneumonia

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STUDIES ON AN UNCOMPLICATED CORYZA OF THE DOMESTIC FOWL

VI. COCCOBACILLIFORM BODIES IN BIRDS INFECTED WITH THE CORYZA OF SLOW ONSET

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PLATE 38

(Received for publication, January 2, 1936)



Introduction

- Mycoplasmas are small prokaryotic organisms
- Without cell wall
- Host specific
- Affinity for mucosal surfaces
- Complex nutritional requirements (difficult to culture)



Typical morphology Mycoplasma spp ('fried egg' morphology)



Period	Subject
1936-1965	Discovery important mycoplasma species
1962-1989	Multifactorial disease (synergism, climate, immunesuppression)
1954-1987	Transmission routes
1956-1974	First control programmes
1956-1965	Antibiotic use & its limitations
1970-1994	Immunisation studies commercial poultry
1955-1994	Diagnostic tests: from serology to molecular analysis

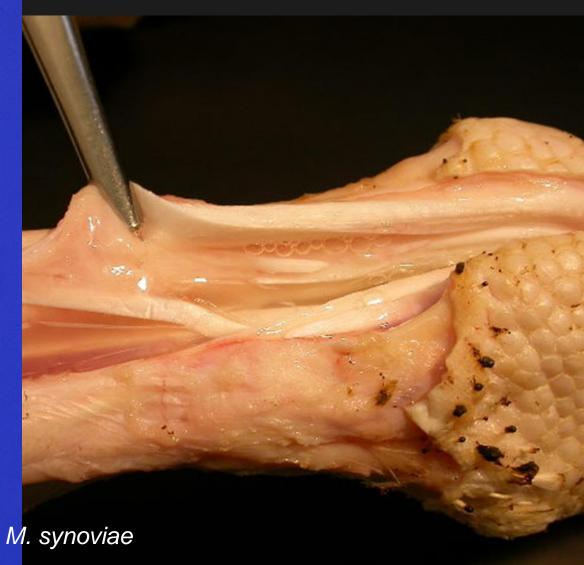


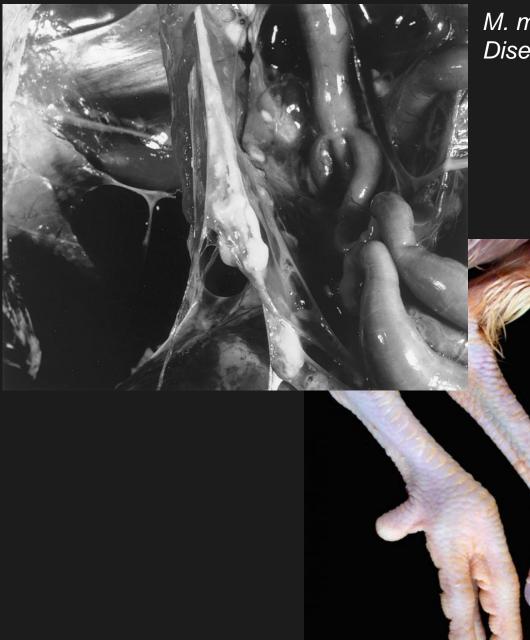
Mycoplasma gallisepticum











A

M. meleagridis Diseases of Poultry (R. Yamamoto)

M. iowae Ley et al., 2010



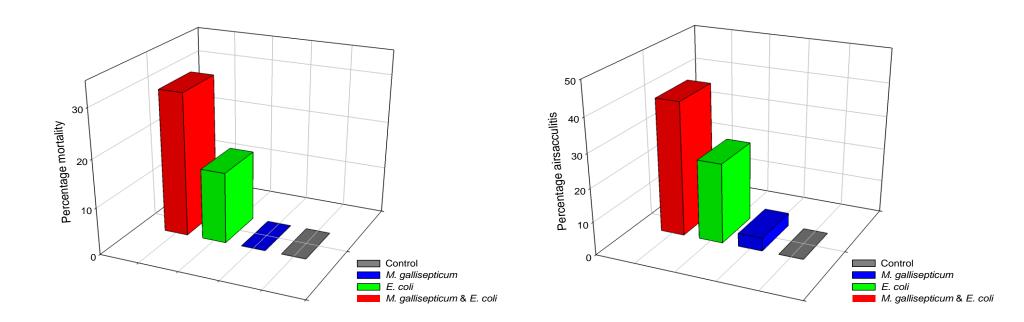


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1-day-old broilers





Bradbury, 2005. Britisch Poult Sci, 46, 125-136

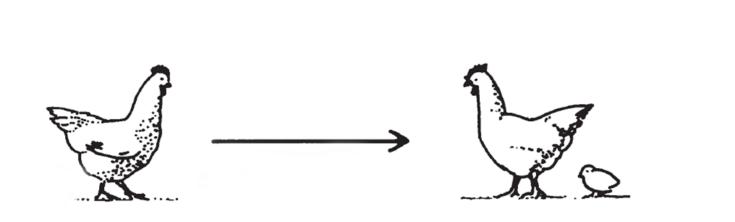
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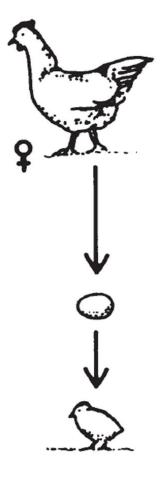


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Transmission routes







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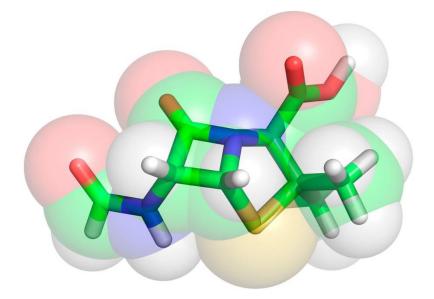






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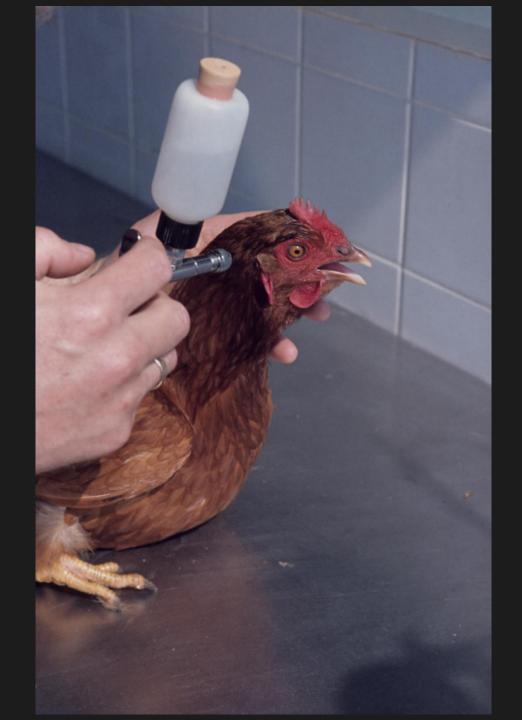




- Temporary effect
- Resistance
- Residues

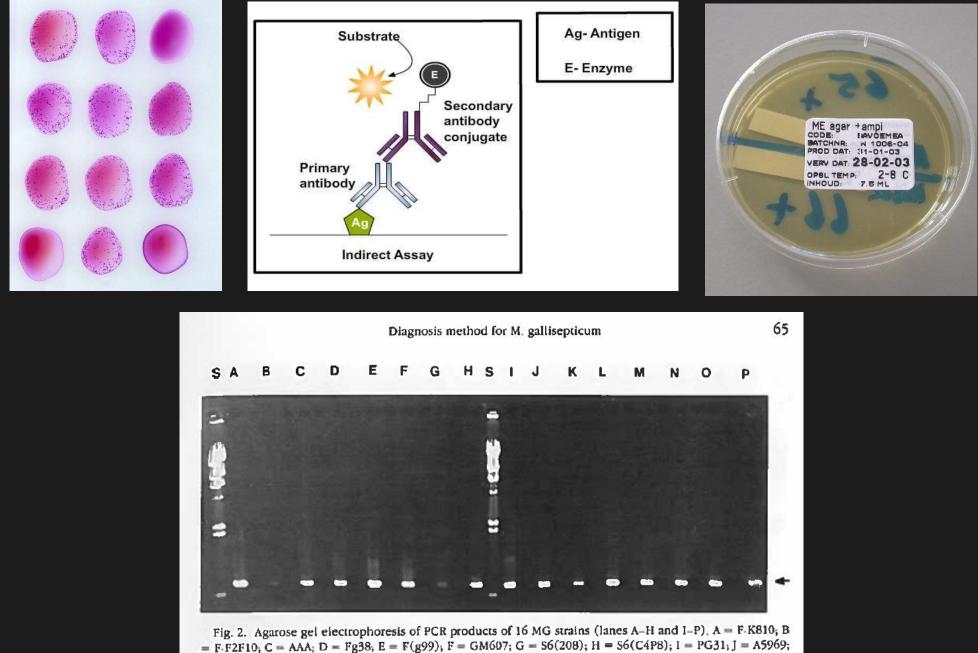


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= F-F2F10; C = AAA; D = Fg38; E = F(g99); F = GM607; G = S6(208); H = S6(C4P8); I = PG31; J = A5969; K = GM747; L = K2101; M = K2101 (36P); N = R; O = V503; and P = F-Conn. Lanes marked with S represent molecular-weight standards (*Hin*dIII-cleaved lambda phage). The arrow corresponds to the 732-bp amplified MG DNA.



Present (90's until now)

Period	Subject
≥1993	Other spp.& strains identified
≥1994	Understanding mycoplasma survival
≥1994	Mycoplasma disease in wild birds
≥1988	Advances in diagnostics
≥1996	Vaccines as control measure
2017	Current situation



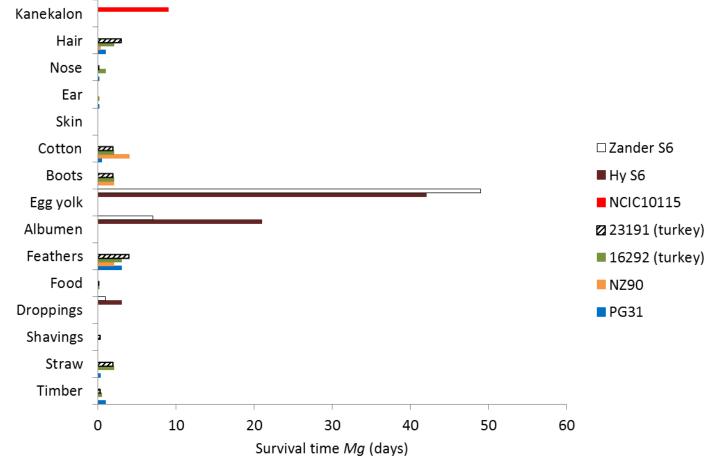
M. synoviae



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Chandiramani et al., 1966; Christensen et al., 1994; Abolink & Gouws, 2014 © GD Animal Health



Veterinary Microbiology 161 (2012) 96-103



Identification of biofilm formation by Mycoplasma gallisepticum

Hongjun Chen¹, Shengqing Yu¹, Meirong Hu, Xiangan Han, Danqing Chen, Xusheng Qiu, Chan Ding^{*}

Shanghai Veterinary Research Institute, Chinese Academy of Agricultural Sciences, 518 Ziyue Road, Shanghai 200241, PR China

- Variation between strains
- Biofilm forming *M. gallisepticum* strains seems more resistant to antibiotic treatment and desinfectance
- Suvival advantage outside the host © GD Animal Health





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House finch (*Carpadocus mexicanus*)

American goldfinch (*Spinus tristis*)

1^{ary} host, 60% †

2^{ary} host



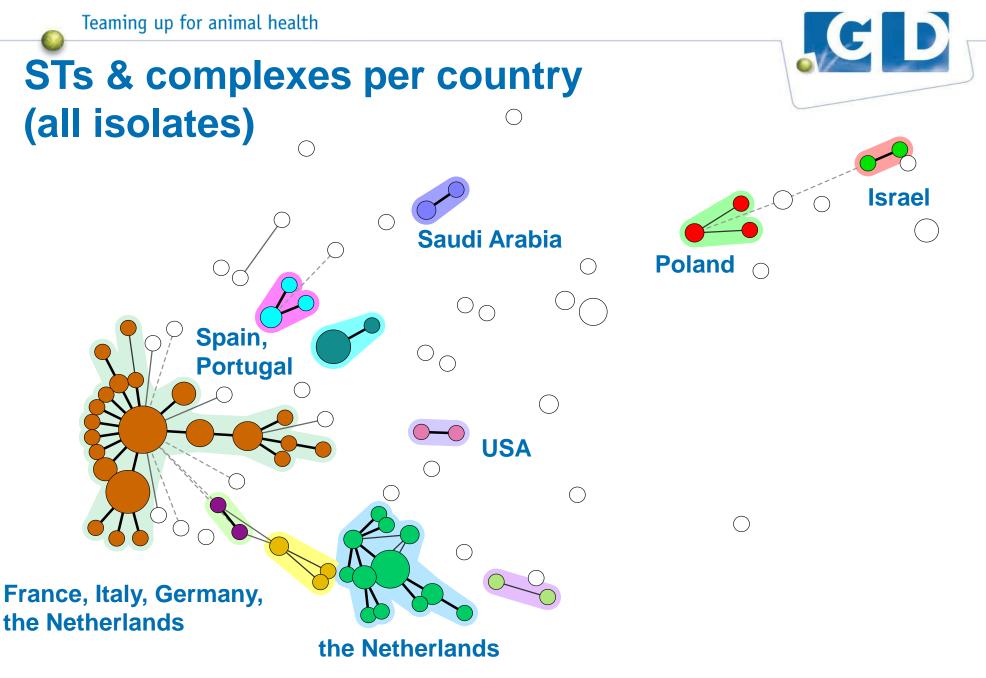
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Molecular typing techniques

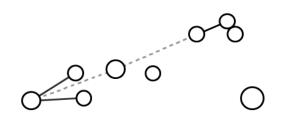
- Band-based
 - PFGE-RAPD-AFLP
- Sequence-based
 - Single gene (vlhA typing)
 - More genes (MLST)
 - Whole genome

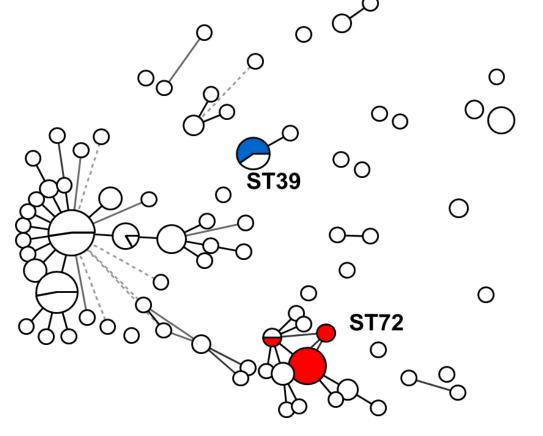


Ο

Ms outbreak







0

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- Farm V (*n* = 8) 09/2003 ST72
- Farm E (*n* = 3) 10/2003 ST72
- Farm W (n = 1) 10/2003 ST72
- Farm M (*n* = 1) 03/2004 ST39
- Farm V (*n* = 2) 10/2004 ST72
- Farm S (*n* = 5) 11/2004 ST39
- Farm F (*n* = 2) 06/2005 ST72

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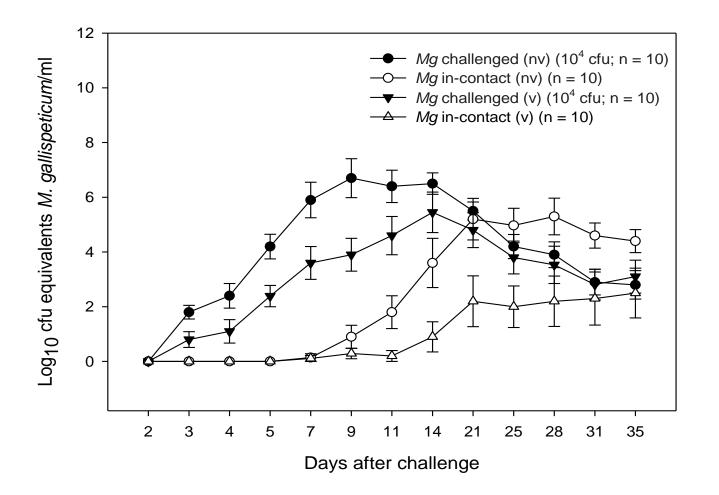
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Effect MS-H vaccine

Shedding (cfu eq./g trachea mucus)

Transmission rate β

8.3 x 10⁶

 1.0×10^{6}

0.022 (0.015-0.031)

0.0012 (0.00048-0.0024)



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Current situation

- Low prevalence *M. gallisepticum* & *M. meleagridis*
 - Control programmes
 - Primary breeding stock free
- High prevalence *M. synoviae*
 - Primary breeding stock free
- M. iowae
 - sporadically reported in commercial poultry

Future

- Reduction prevalence
 - M. gallisepticum developing countries
 - M. synoviae worldwide
- Shift to molecular-based tests
 - Earlier/accurate detection infections
- Prudent use antibiotics (MIC)
- Whole genome sequence
- DIVA tests
- Biosecurity







Thank you for your attention

