

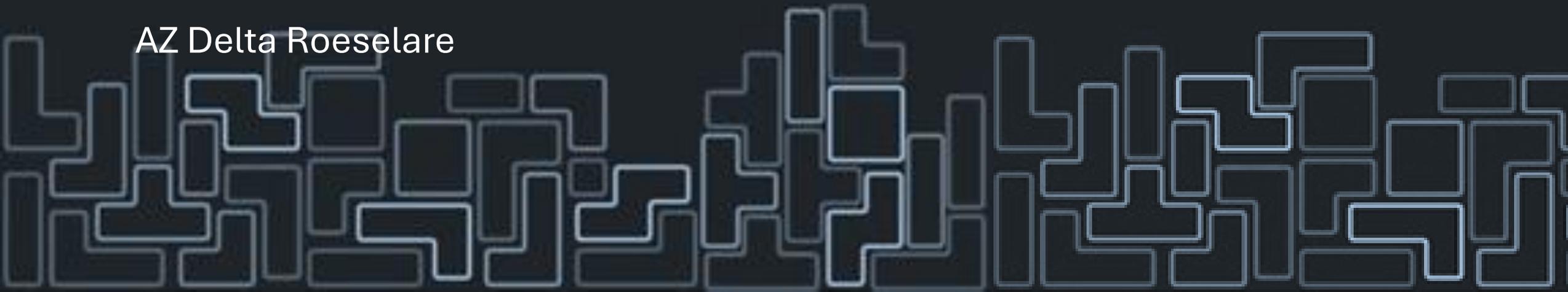
# New autoantibodies

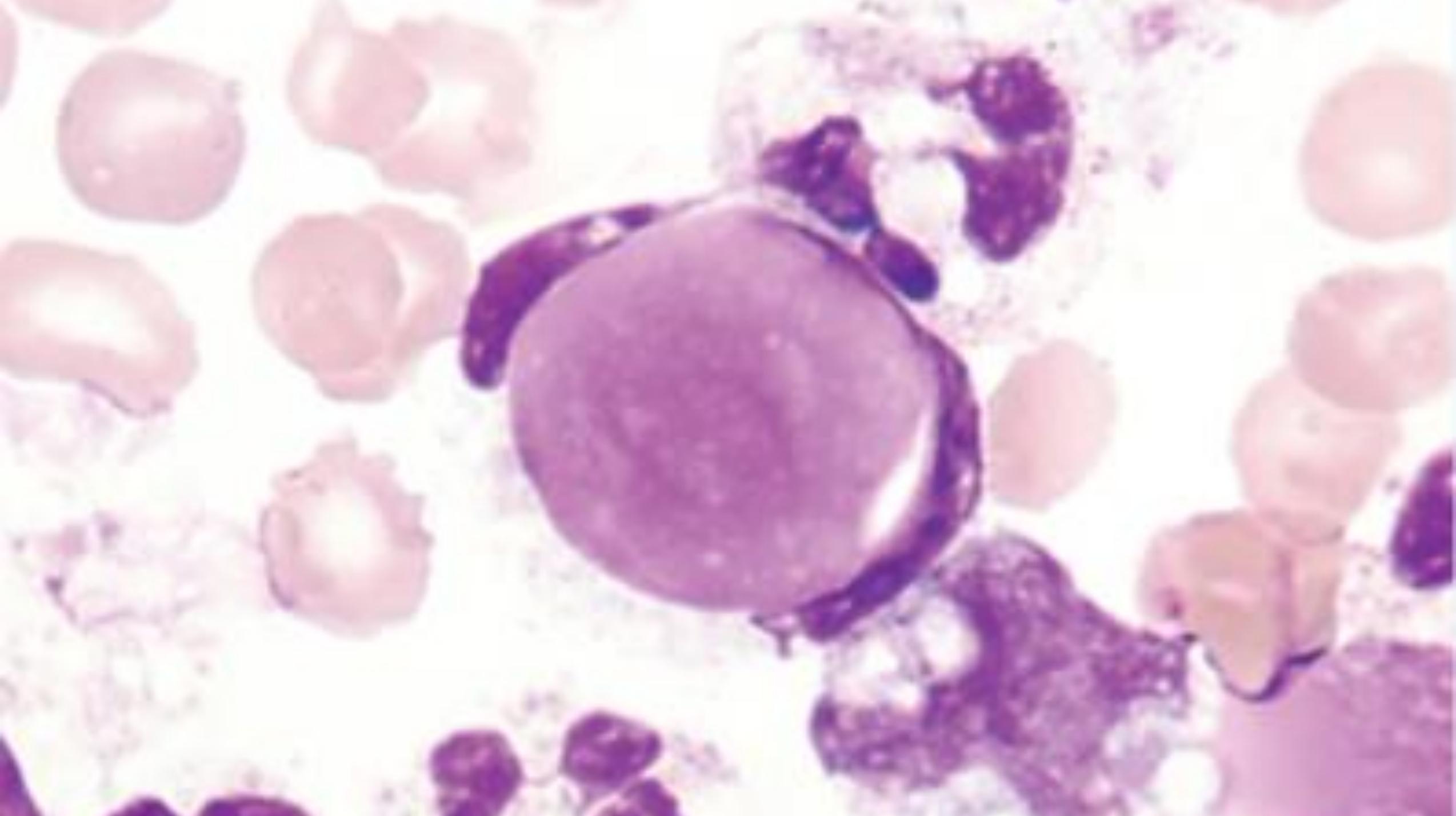
and the role of **IP-MS** in routine diagnostics ?

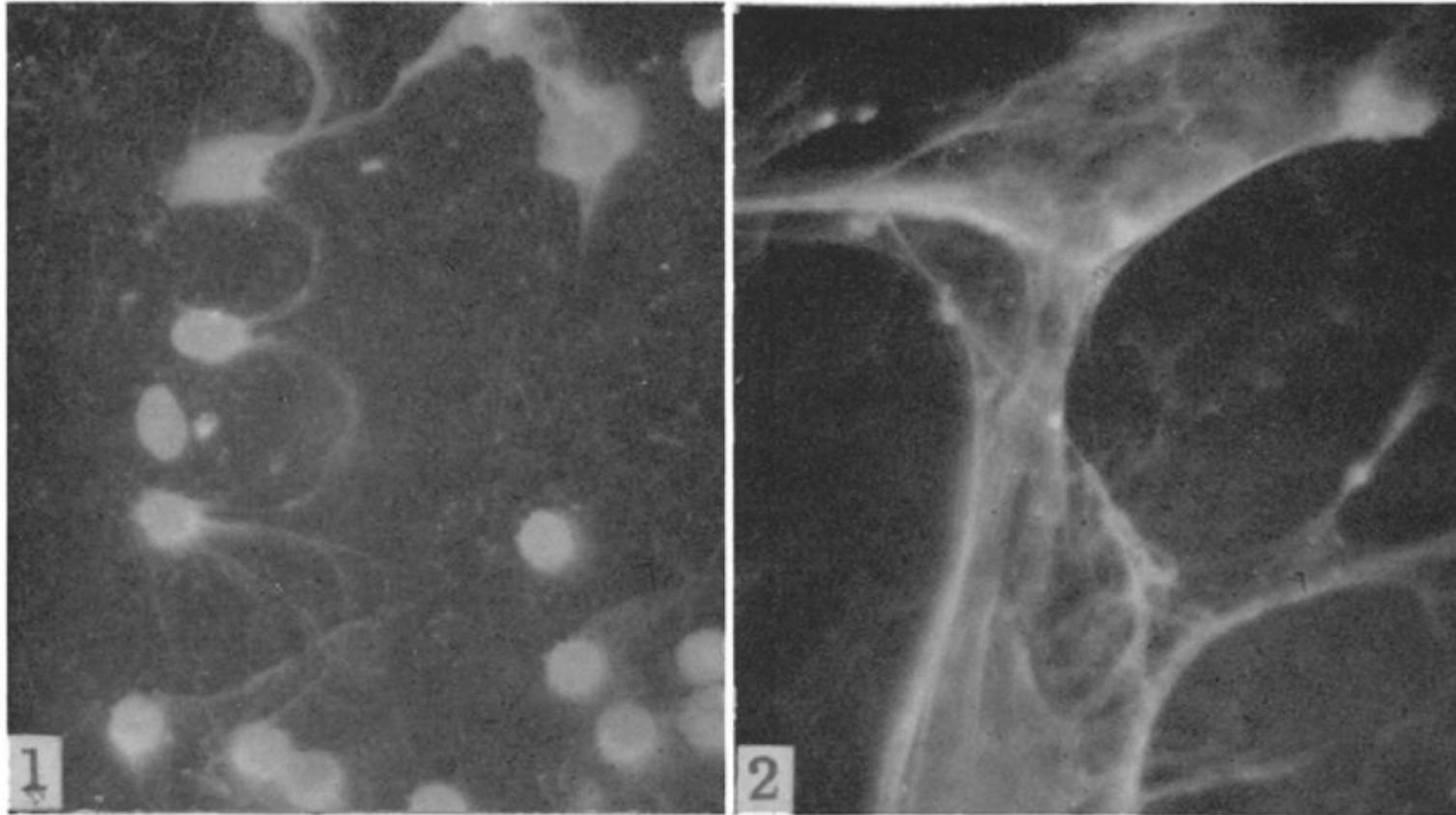
Jean-Baptiste Vulsteke MD PhD

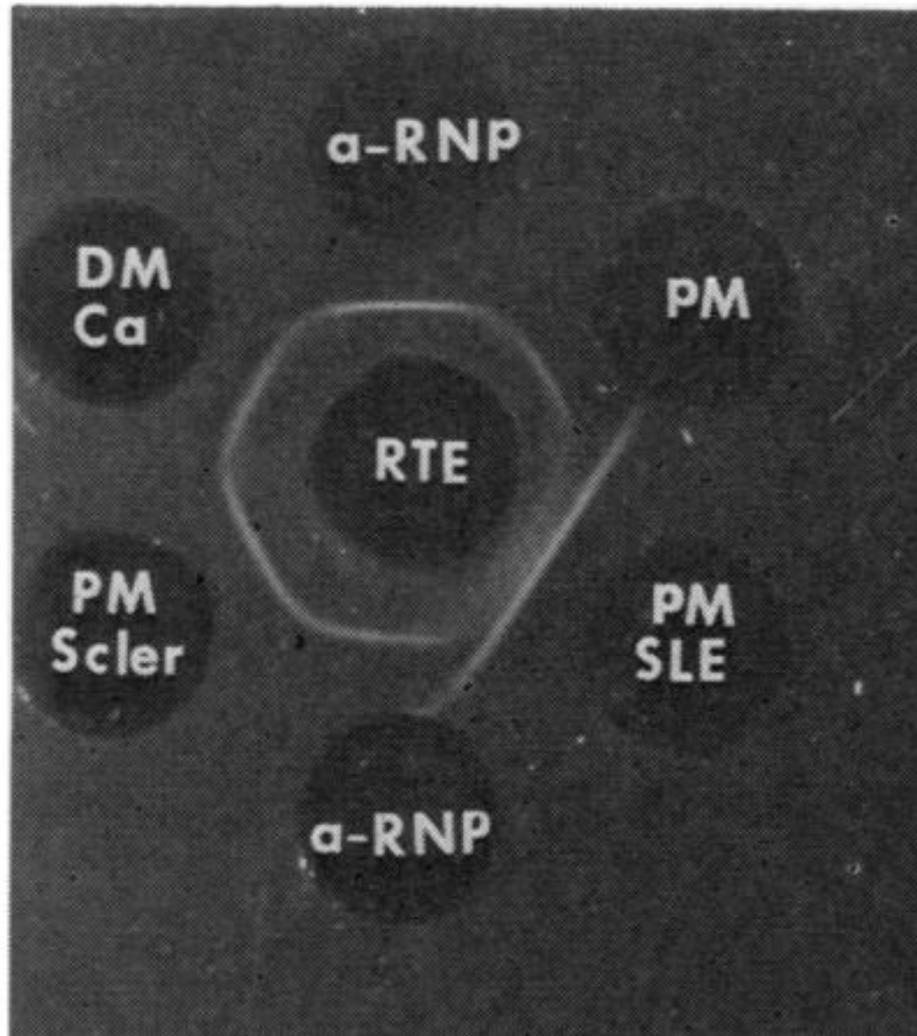
University Hospitals Leuven

AZ Delta Roeselare

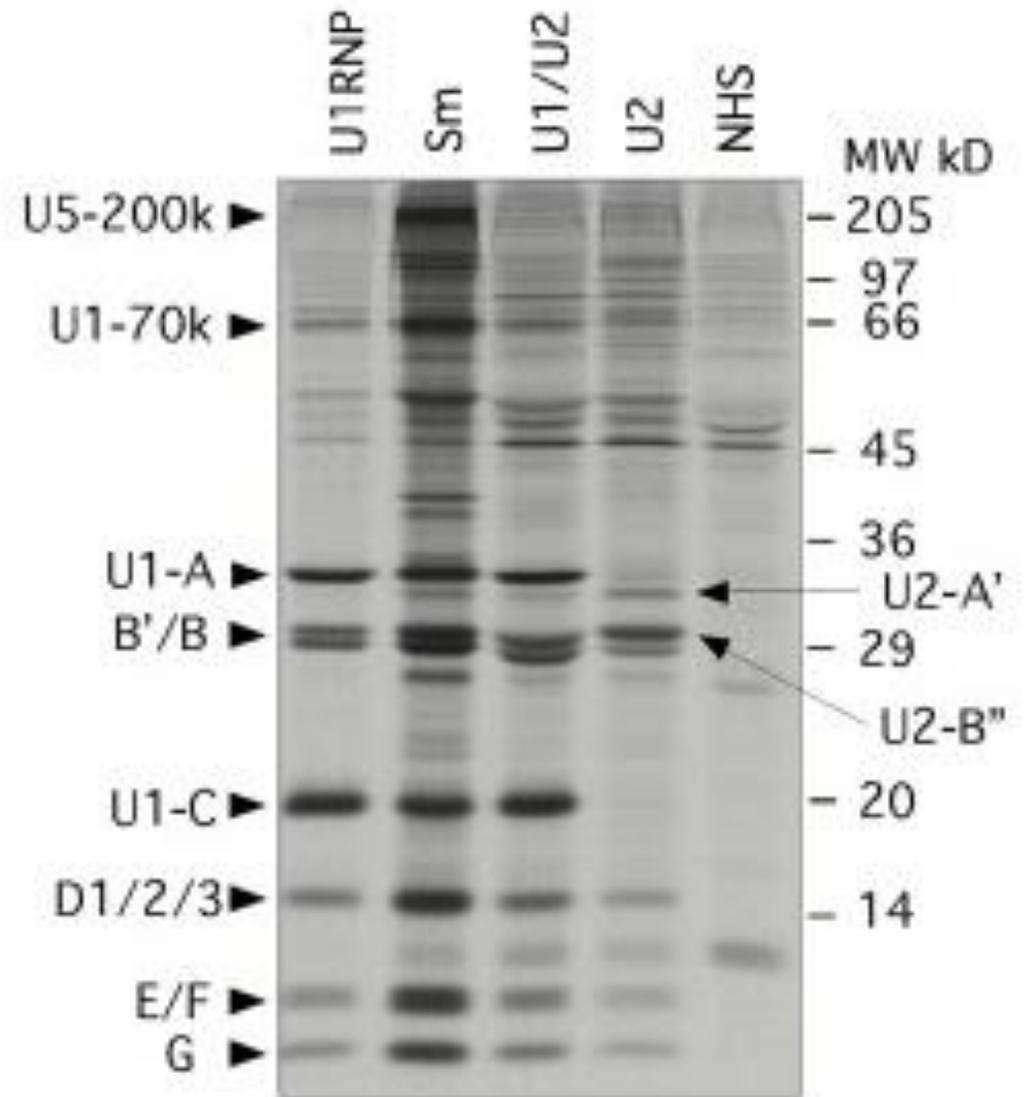




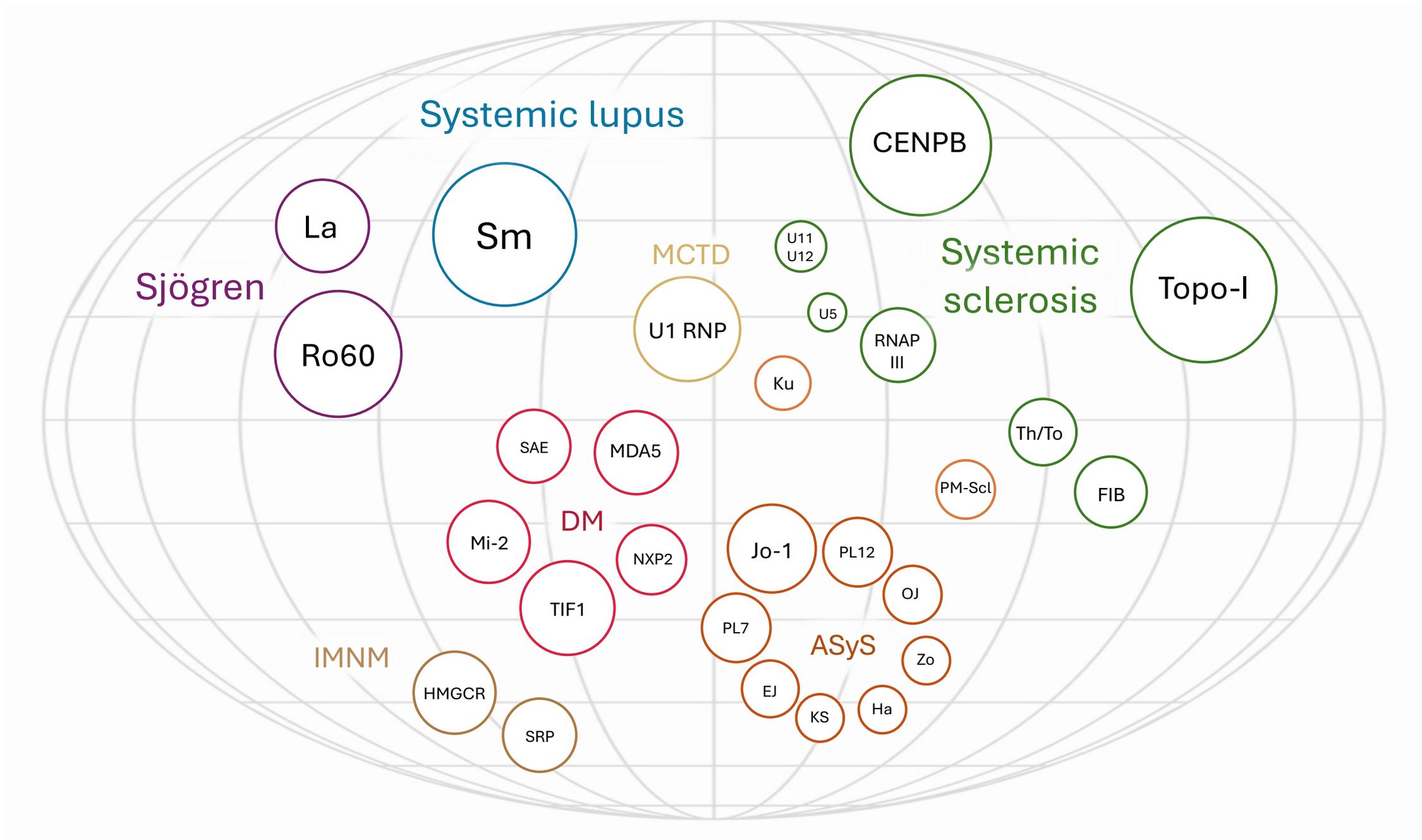




Venables et al., ARD 1981



Sato et al., Autoantibodies 2014



Systemic lupus

CENPB

La

Sm

MCTD

U11  
U12

Systemic  
sclerosis

Topo-I

Sjögren

Ro60

U1 RNP

U5

RNAP  
III

Ku

SAE

MDA5

Th/To

PM-Scl

FIB

DM

Mi-2

NXP2

Jo-1

PL12

OJ

TIF1

PL7

ASyS

Zo

IMNM

HMGCR

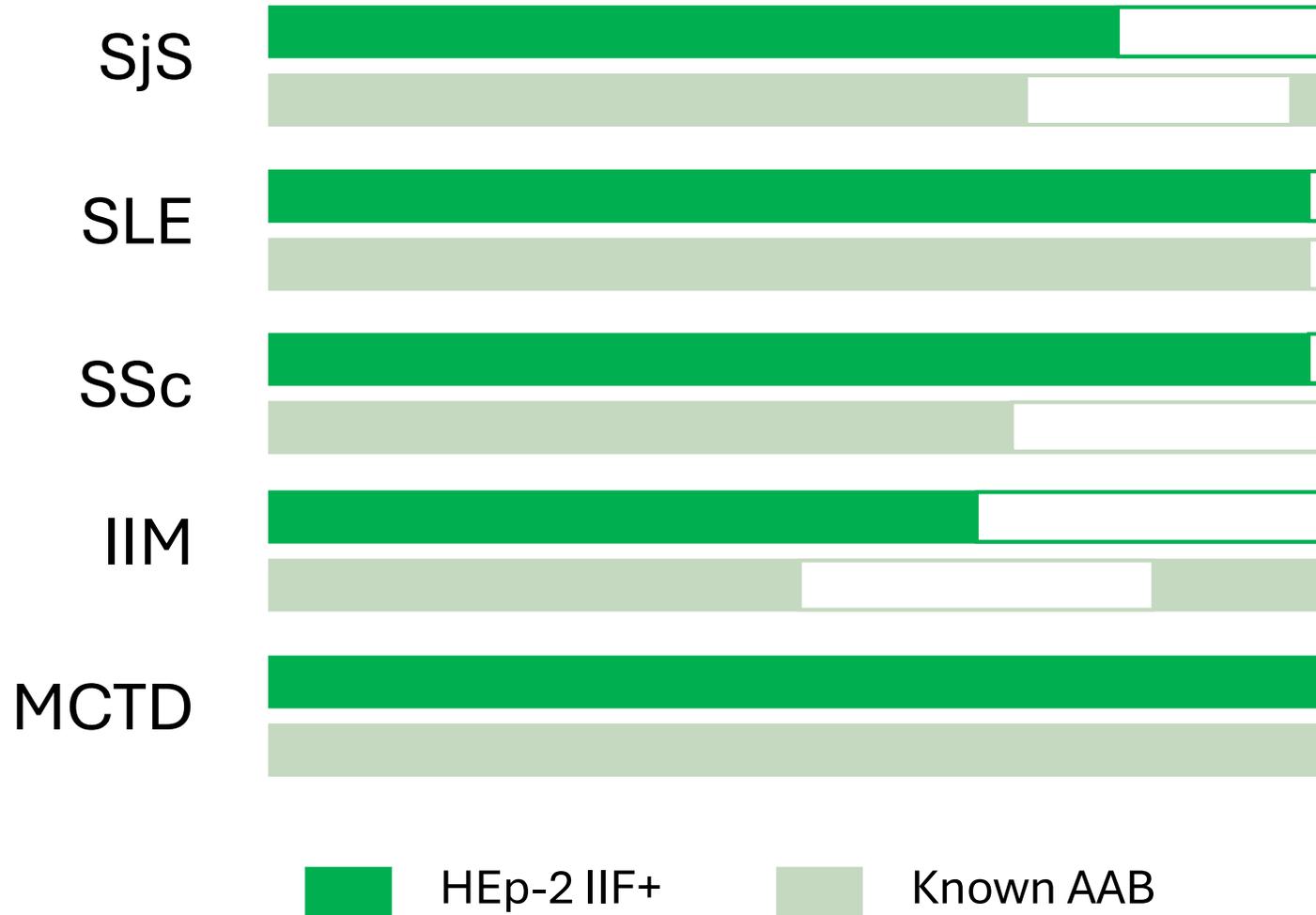
EJ

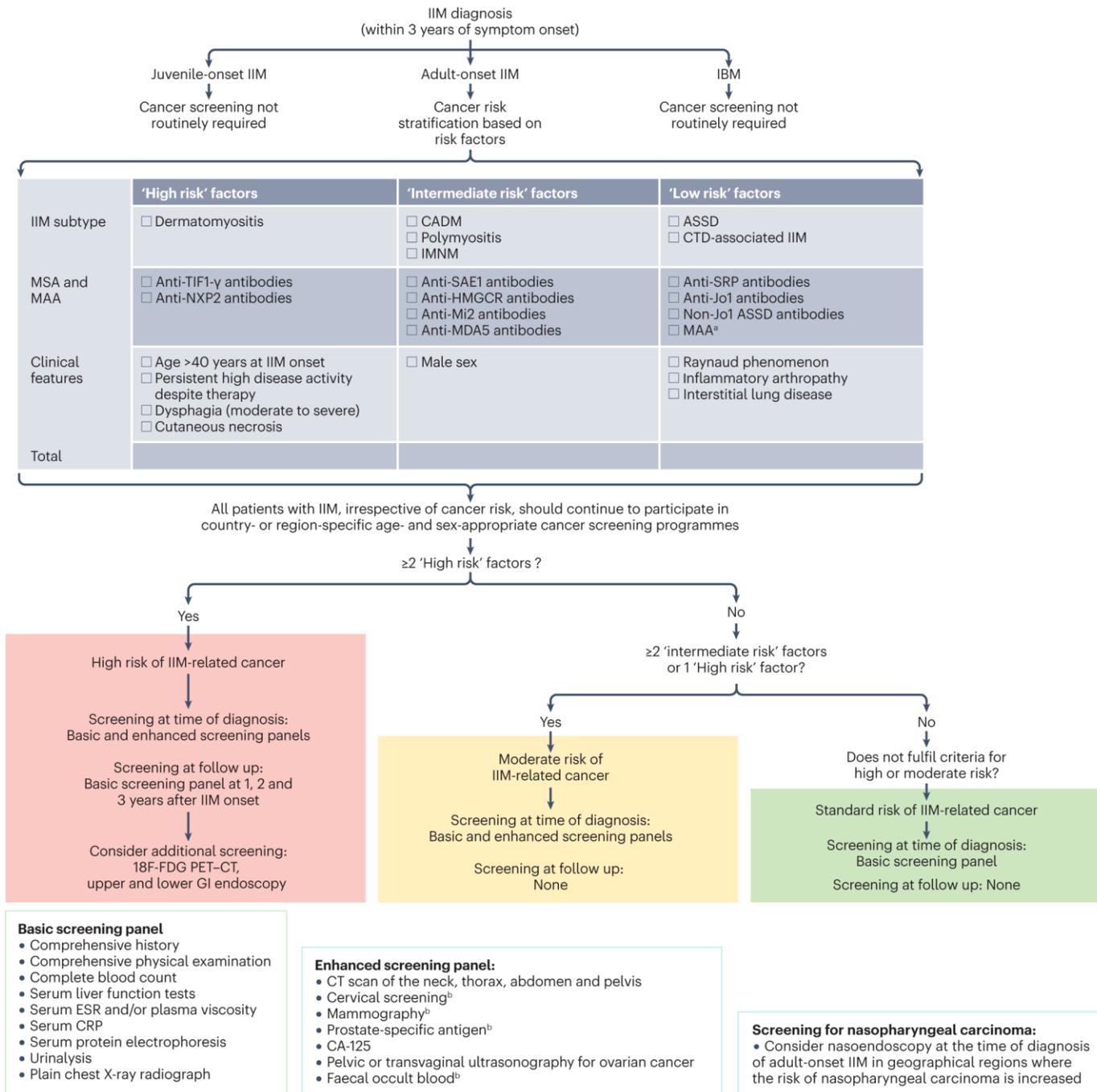
KS

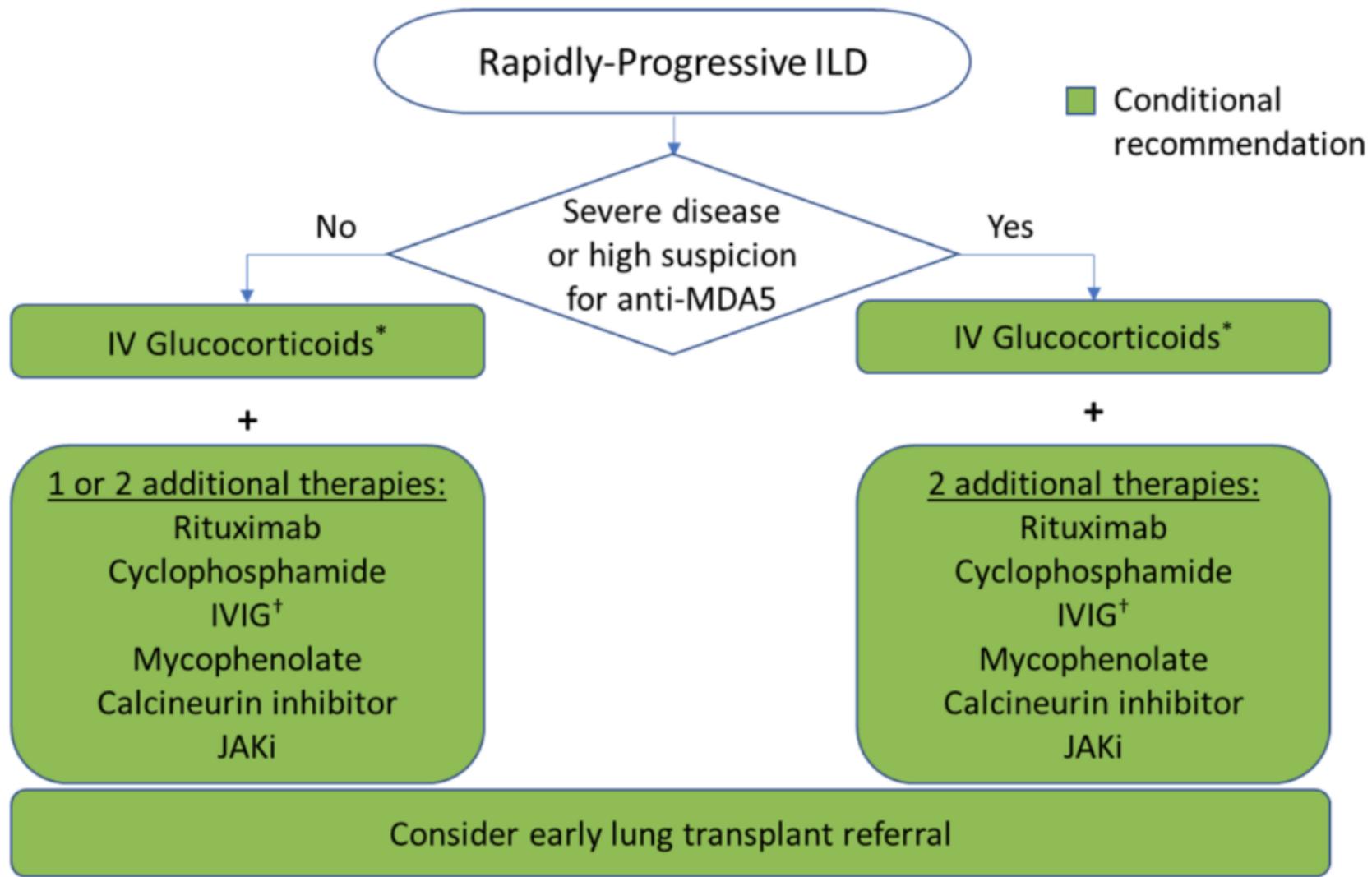
Ha

SRP

# Anno 2024: a seronegative gap remains





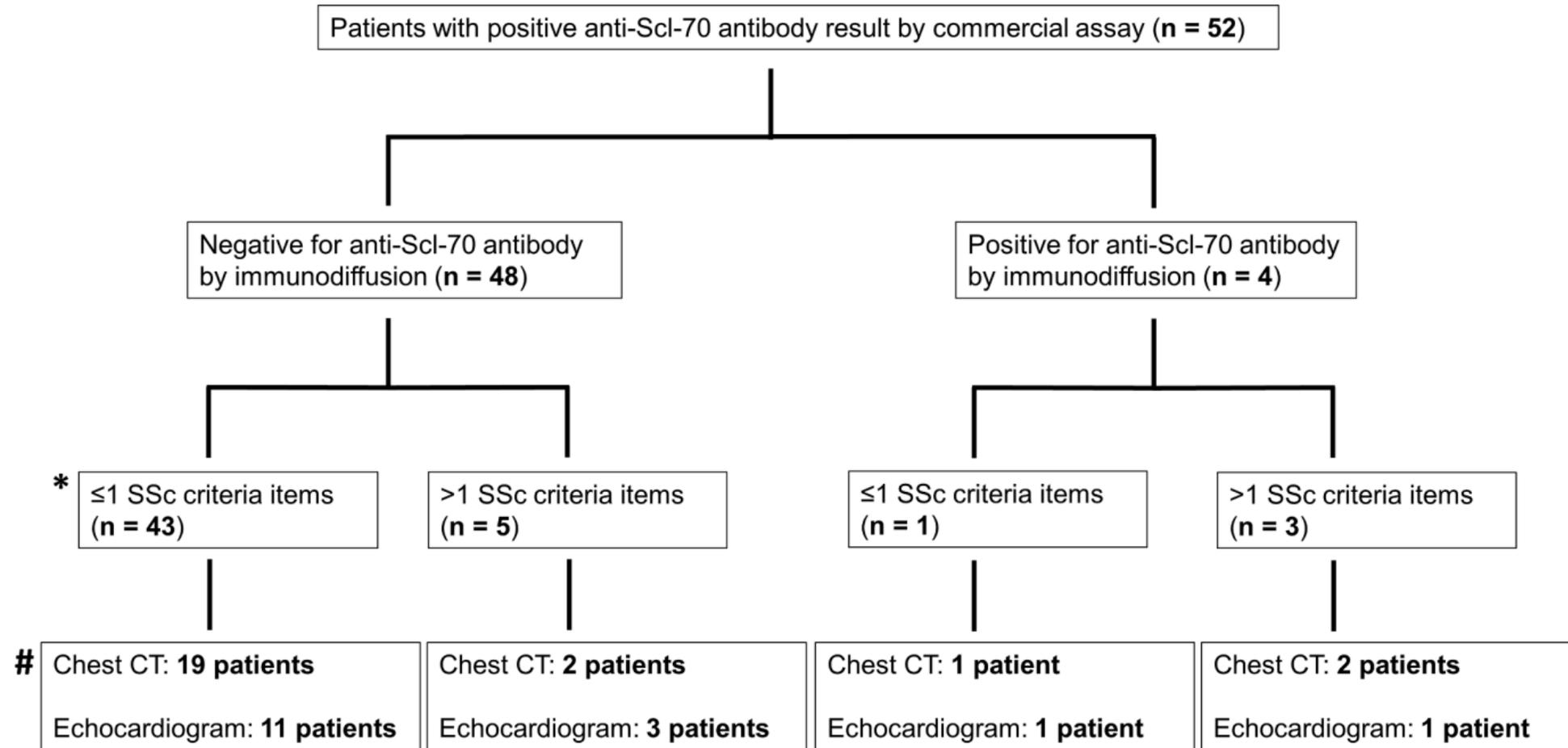


# Anno 2024: a brewing solid-phase storm

- **Discordance between solid-phase assays** Vulsteke et al., Ann Rheum Dis 2018
- **Discordance solid-phase assays and immunoprecipitation**
- **Lower pretest probabilities**

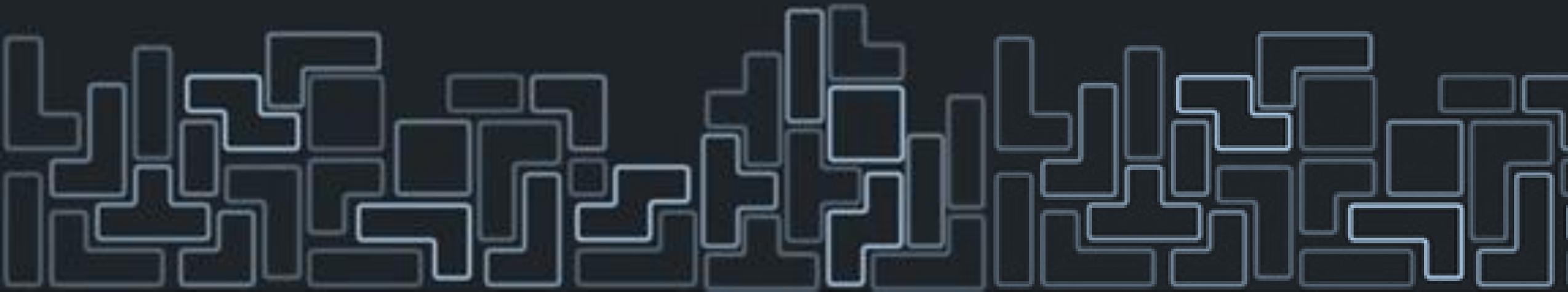
<b>Table 1</b> Agreement between LIA and IP derived from three studies					
	<i>Espinosa-Ortega et al</i>	<i>Cavazzana et al</i>	<i>Mahler et al</i>	<b>Combined</b>	
<b>MSA</b>	<b>DP/IP/LP/DN</b> <b>Kappa (95% CI)</b>				<b>Interpretation (kappa)</b>
Jo-1	11/1/7/91 0.69 (0.50 to 0.89)	3/5/7/42 0.21 (−0.11 to 0.52)	NA	14/6/14/133 0.52 (0.33 to 0.70)	Minimal–Moderate
TIF1y	1/2/0/107 0.56 (0.29 to 0.83)	4/0/2/51 0.78 (0.49 to 1.00)	11/4/4/138 0.71 (0.51 to 0.90)	16/6/6/296 0.71 (0.55 to 0.86)	Weak–Strong
SRP	3/1/0/106 0.85 (0.57 to 1.00)	0/3/7/47 −0.08 (−0.15 to −0.01)	14/1/8/134 0.73 (0.56 to 0.89)	17/5/15/287 0.60 (0.44 to 0.76)	Weak–Moderate
MDA5	1/2/0/107 0.49 (−0.12 to 1.00)	3/1/2/51 0.64 (0.26 to 1.00)	14/1/5/137 0.80 (0.65 to 0.96)	18/4/7/295 0.75 (0.61 to 0.89)	Moderate–Strong
PM/Scl	2/3/1/104 0.48 (0.05 to 0.92)	NA NA	NA NA	2/3/1/104 0.48 (0.05 to 0.92)	NA
Ku	3/0/1/106 0.86 (0.57 to 1.00)	NA NA	NA NA	3/0/1/106 0.86 (0.57 to 1.00)	NA
SAE	3/0/1/106 0.86 (0.57 to 1.00)	0/2/1/54 −0.02 (−0.06 to 0.01)	15/0/3/139 0.90 (0.79 to 1.00)	18/2/5/299 0.83 (0.70 to 0.95)	Moderate–Almost perfect
NXP2	0/0/2/108 0.00	5/3/2/47 −0.62 (0.31 to 0.92)	13/2/0/142 0.92 (0.81 to 1.00)	18/5/4/297 0.79 (0.65 to 0.92)	Moderate–Almost perfect
Mi-2	2/0/6/102 0.38 (0.00 to 0.76)	3/0/5/49 0.51 (0.45 to 0.87)	12/3/9/133 0.62 (0.43 to 0.82)	17/3/20/284 0.56 (0.41 to 0.72)	Weak–Moderate
EJ	0/0/0/110 NA	1/2/0/54 0.49 (−0.11 to 1.00)	7/3/1/146 0.76 (0.54 to 0.99)	8/5/1/310 0.72 (0.50 to 0.93)	Weak–Almost perfect
OJ	0/1/0/109 0.00	0/2/0/55 0.00	NA	0/3/0/164 NA	NA
PL-7	0/0/0/110 NA	0/0/2/55 0.00	12/3/0/142 0.88 (0.74 to 1.00)	12/3/2/307 0.82 (0.67 to 0.97)	Moderate–Almost perfect
PL-12	0/0/1/109 0.00	0/0/3/54 0.00	14/1/5/137 0.80 (0.65 to 0.96)	14/1/9/300 0.72 (0.56 to 0.89)	Weak–Strong

95% CIs were reconstructed based on the given prevalence.

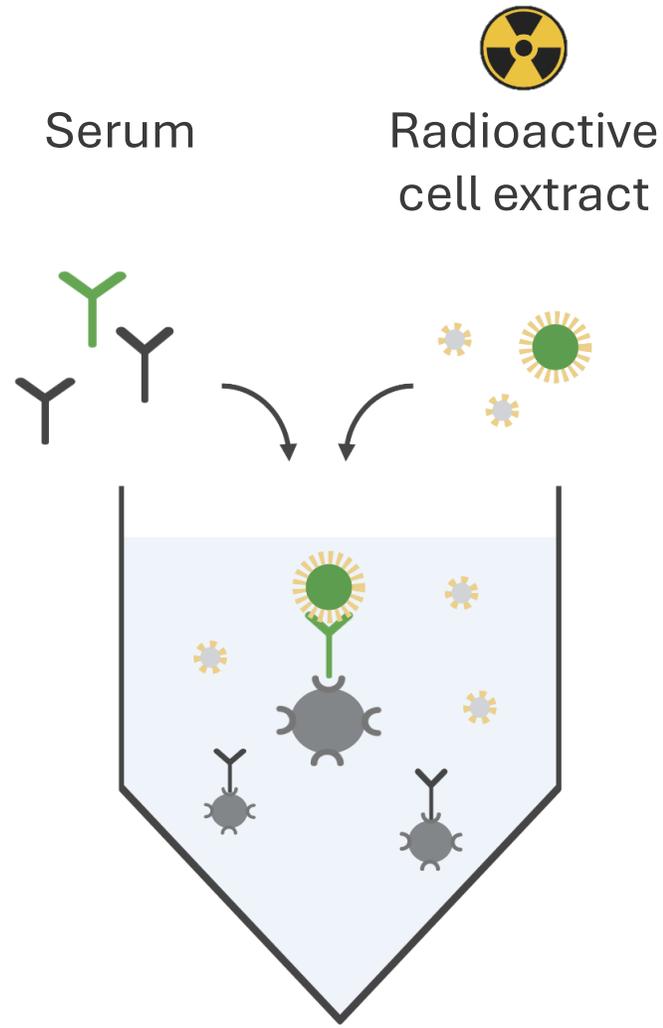


**Fig. 2.** Flow diagram showing numbers of patients who underwent chest CT and/or echocardiogram within patient subgroups, by Scl-70 immunodiffusion results and number of ACR/EULAR systemic sclerosis classification criteria items. \*Commercial Scl-70 antibody results were not counted towards the criteria items, as noted in the Methods section. #Refers to the numbers of patients who underwent chest CT and/or echocardiogram prior to evaluation in our clinic.

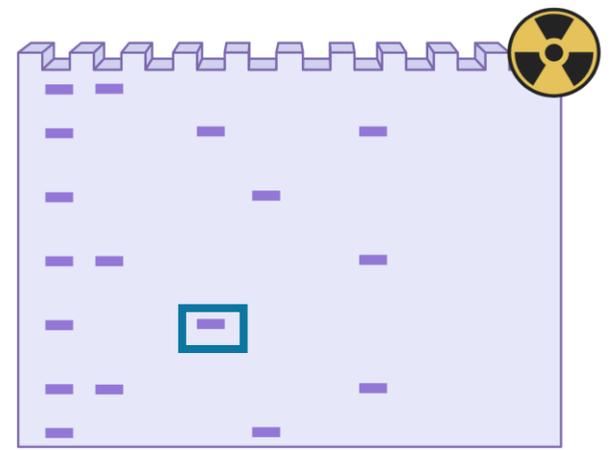
# **Immunoprecipitation-mass spectrometry or IP-MS**



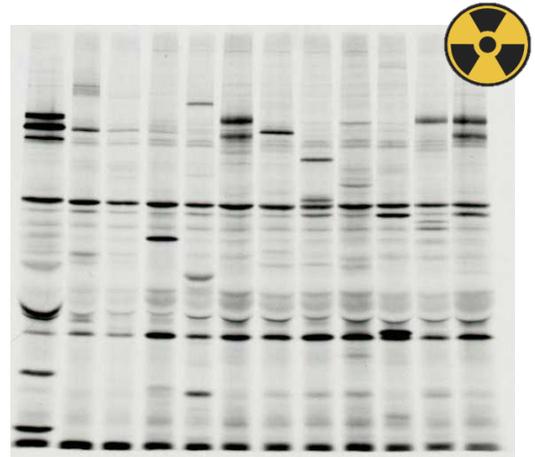
# Immunoprecipitation



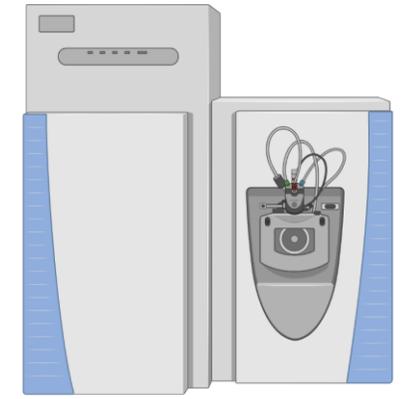
# Gel electrophoresis



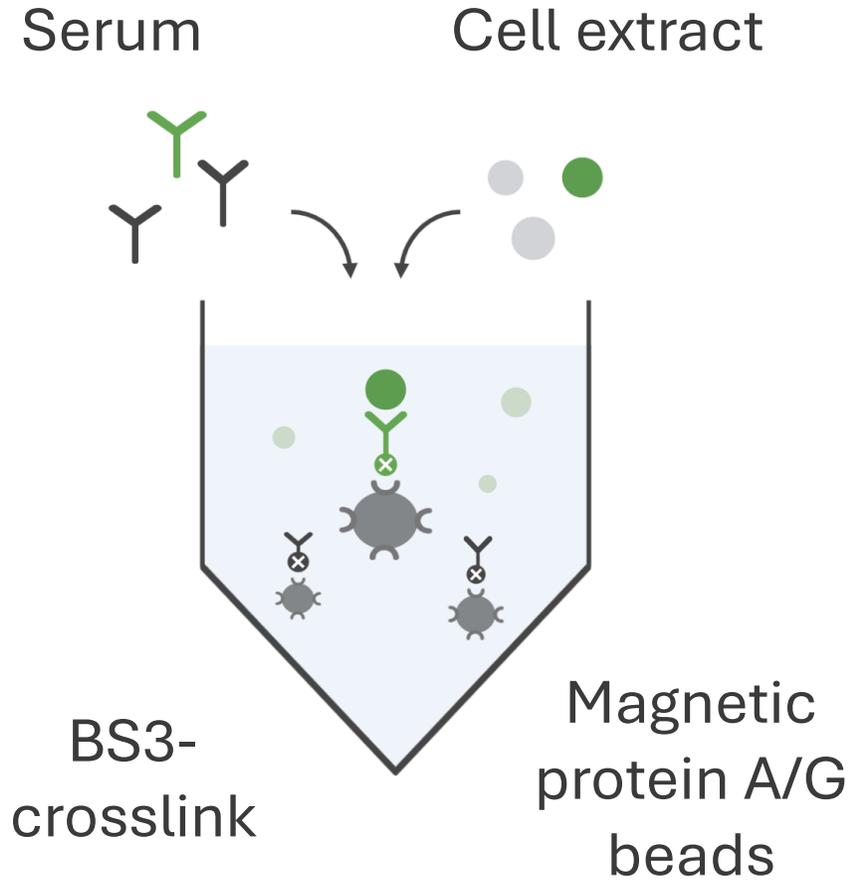
# Autoradiography



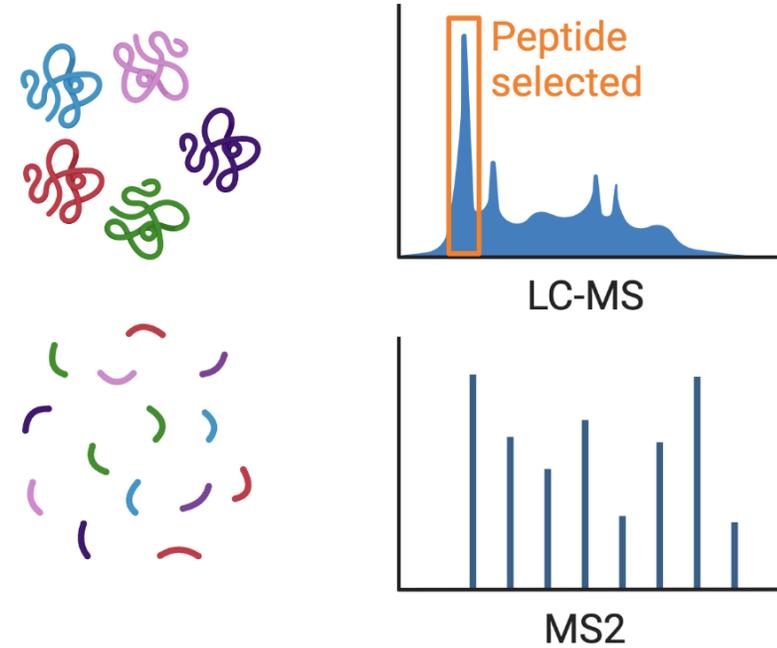
# Mass spectrometry



# Immunoprecipitation



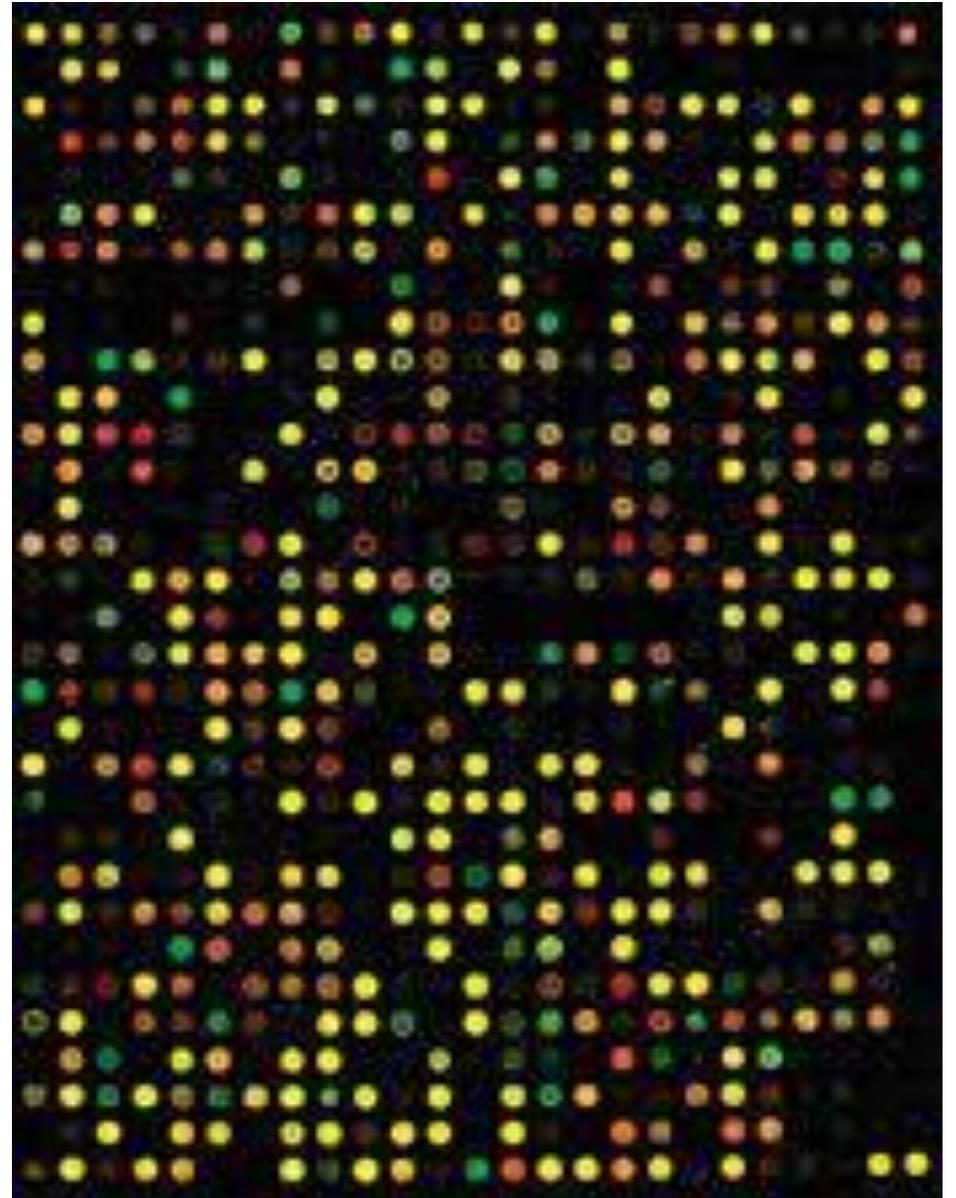
# LC-MS/MS



Data-dependent acquisition

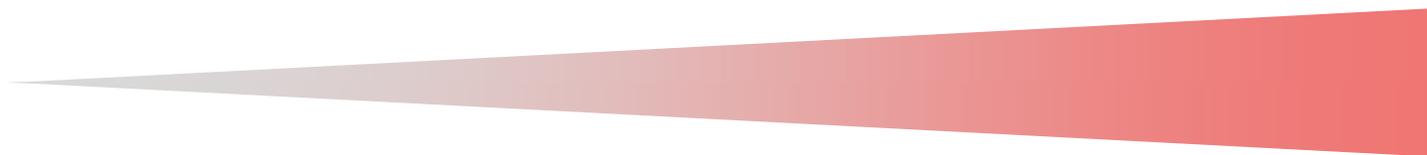
# Why not protein arrays?

- None of the established AABs have been identified with protein arrays
- Conformational dependence?





Quaternary  
autoepitopes?



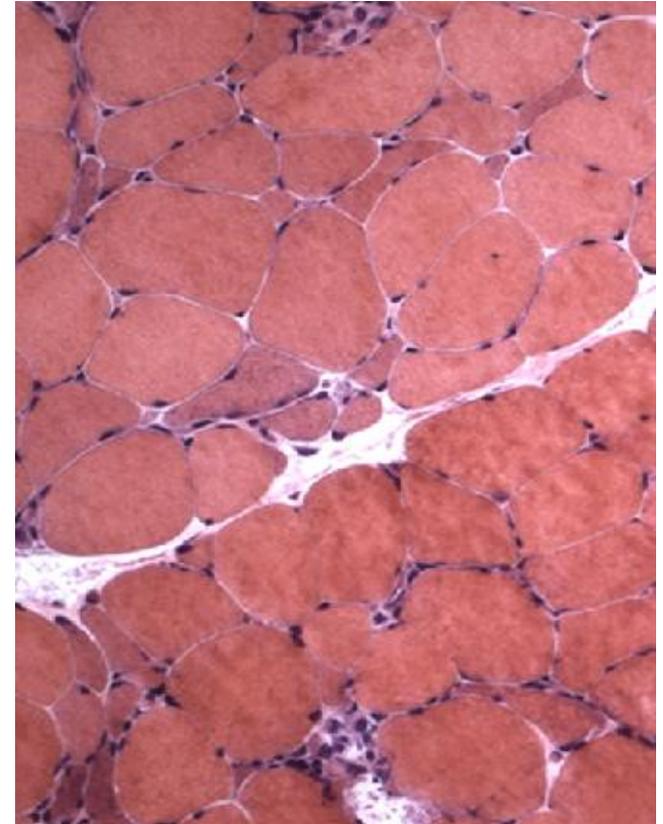
Jo-1      Ro60

PM-Scl      OJ  
RuvBL

Heterogeneity

Mr. N

72 y



Mr. N

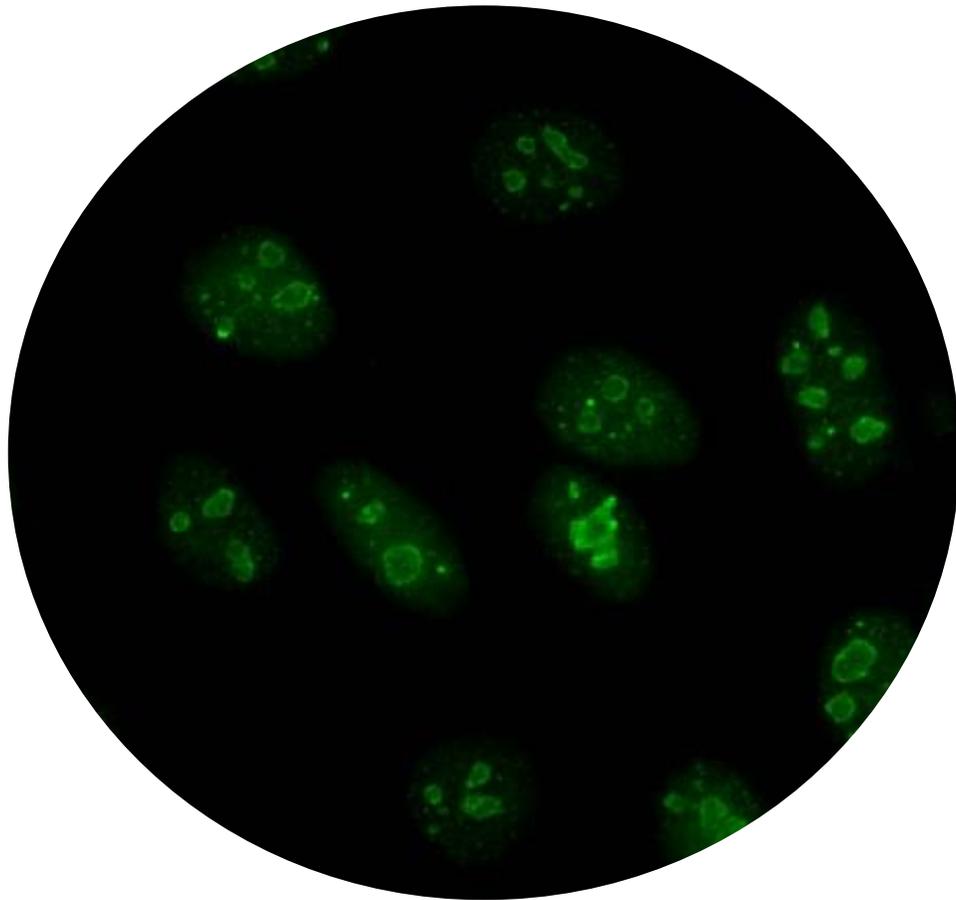


Cluster of Vimentin	VIM	369	314	314	167	303	70	127	88	84	92	105	85	71	91	145	90	127
RNA transcription, translation and transport factor protein	RTRAF	105	103	94	97	84	66	71	55	13	63	78	60	32	83	76	28	70
Signal recognition particle 14 kDa protein	SRP14	66	57	66	46	56	49	41	44	20	39	46	25	17	38	48	51	37
Cluster of Actin, cytoplasmic 1	ACTB	65	57	64	51	43	20	38	24	27	28	44	22	13	28	37	25	39
Pre-mRNA-processing factor 19	PRPF19	50	32	61	44	39	38	16	20	5	14	27	16	15	40	25	12	58
Non-POU domain-containing octamer-binding protein	NONO	59	71	61	52	70	29	28	16	15	42	35	12	22	19	29	52	48
Cluster of Prohibitin	PHB	66	48	60	30	52	42	33	30	17	31	47	27	17	40	50	25	39
40S ribosomal protein S18	RPS18	69	56	54	47	56	25	36	26	25	41	49	21	24	25	39	35	33
Cleavage and polyadenylation specificity factor subunit 5	NUDT21	50	54	49	21	49	33	31	30	18	27	35	18	7	32	32	13	32
Cluster of Heat shock cognate 71 kDa protein	HSPA8	42	40	46	28	36	21	21	18	23	14	21	20	24	29	11	16	23
Splicing factor, proline- and glutamine-rich	SFPQ	44	46	44	44	42	32	25	22	31	32	24	31	34	40	39	30	35
Cluster of Uncharacterized protein DKFZp686N02209		13	12	44	59	140	63	43	48	61	67	67	103	56	52	90	13	136
DNA damage-binding protein 1	DDB1	56	48	43	36	34	31	18	33	16	25	19	19	26	29	20	8	27
Cluster of Polyadenylate-binding protein 1	PABPC1	43	36	42	18	33	15	15	16	1	15	20	14	8	23	21	5	10
Prohibitin-2	PHB2	41	31	37	19	33	29	23	25	24	24	28	23	21	21	23	13	21
Cluster of Heat shock 70 kDa protein 1A	HSPA1A	35	41	34	25	35	28	24	28	28	22	27	28	31	33	17	12	22
40S ribosomal protein S3	RPS3	30	31	32	25	30	33	23	32	16	29	29	16	11	24	22	36	18
Pyruvate kinase PKM	PKM	34	29	31	30	29	42	35	33	13	32	31	27	21	28	31	13	15
40S ribosomal protein S5	RPS5	41	37	29	19	21	22	14	7	2	11	22	5	0	6	28	24	12
Dolichyl-diphosphooligosaccharide--protein glycosyltransferase 48 kDa subunit	DDOST	29	34	29	22	23	14	12	4	4	8	21	0	0	12	16	5	9
Nucleolar and coiled-body phosphoprotein 1	NOLC1	0	0	28	0	0	0	0	0	0	0	0	0	0	0	0	0	0
Cluster of Myosin light polypeptide 6	MYL6	29	21	25	11	17	8	10	7	8	6	9	10	9	12	8	5	7
40S ribosomal protein SA	RPSA	26	26	24	25	24	24	18	22	16	22	24	24	16	24	16	13	34
60S ribosomal protein L13	RPL13	27	30	24	23	27	12	18	7	15	13	13	18	20	18	16	22	4
NADH dehydrogenase [ubiquinone] iron-sulfur protein 3, mitochondrial	NDUFS3	6	14	22	9	18	12	9	13	2	7	13	6	2	10	8	1	5
Ribosomal RNA small subunit methyltransferase NEP1	EMG1	21	17	19	12	15	9	8	9	0	6	10	6	2	15	9	3	9
40S ribosomal protein S7	RPS7	23	10	18	7	9	0	0	1	0	0	2	0	0	1	9	1	2
60S acidic ribosomal protein P2	RPLP2	30	20	18	8	13	7	5	7	6	6	16	15	4	23	13	9	16
Cluster of General transcription and DNA repair factor IIH helicase subunit XPB	ERCC3	1	0	18	0	0	0	0	0	0	0	0	0	0	0	0	0	0
RuvB-like 1	RUVBL1	13	18	17	6	16	4	7	4	1	0	10	1	2	2	2	0	2
60S ribosomal protein L23	RPL23	14	19	17	23	12	11	16	11	13	11	12	12	9	14	9	8	10
Replication factor C subunit 4	RFC4	13	8	17	3	10	5	2	0	2	5	2	6	4	5	4	0	3
Cytochrome c oxidase subunit 7A2, mitochondrial	COX7A2	15	15	17	15	15	7	9	7	0	8	11	10	0	4	10	5	7

Mr. N

Cluster of Vimentin  
 RNA transcription, translation and transport factor protein  
 Signal recognition particle 14 kDa protein  
 Cluster of Actin, cytoplasmic 1  
 Pre-mRNA-processing factor 19  
 Non-POU domain-containing octamer-binding protein  
 Cluster of Prohibitin  
 40S ribosomal protein S18  
 Cleavage and polyadenylation specificity factor subunit 5  
 Cluster of Heat shock cognate 71 kDa protein  
 Splicing factor, proline- and glutamine-rich  
 Cluster of Uncharacterized protein DKFZp686N02209  
 DNA damage-binding protein 1  
 Cluster of Polyadenylate-binding protein 1  
 Prohibitin-2  
 Cluster of Heat shock 70 kDa protein 1A  
 40S ribosomal protein S3  
 Pyruvate kinase PKM  
 40S ribosomal protein S5  
 Dolichyl-diphosphooligosaccharide--protein glycosyltransferase 48 kDa subunit  
 Nucleolar and coiled-body phosphoprotein 1  
 Cluster of Myosin light polypeptide 6  
 40S ribosomal protein SA  
 60S ribosomal protein L13  
 NADH dehydrogenase [ubiquinone] iron-sulfur protein 3, mitochondrial  
 Ribosomal RNA small subunit methyltransferase NEP1  
 40S ribosomal protein S7  
 60S acidic ribosomal protein P2  
 Cluster of General transcription and DNA repair factor IIH helicase subunit XPB  
 RuvB-like 1  
 60S ribosomal protein L23  
 Replication factor C subunit 4  
 Cytochrome c oxidase subunit 7A2, mitochondrial

VIM	369	314	314	167	303	70	127	88	84	92	105	85	71	91	145	90	127
RTRAF	105	103	94	97	84	66	71	55	13	63	78	60	32	83	76	28	70
SRP14	66	57	66	46	56	49	41	44	20	39	46	25	17	38	48	51	37
ACTB	65	57	64	51	43	20	38	24	27	28	44	22	13	28	37	25	39
PRPF19	50	32	61	44	39	38	16	20	5	14	27	16	15	40	25	12	58
NONO	59	71	61	52	70	29	28	16	15	42	35	12	22	19	29	52	48
PHB	66	48	60	30	52	42	33	30	17	31	47	27	17	40	50	25	39
RPS18	69	56	54	47	56	25	36	26	25	41	49	21	24	25	39	35	33
NUDT21	50	54	49	21	49	33	31	30	18	27	35	18	7	32	32	13	32
HSPA8	42	40	46	28	36	21	21	18	23	14	21	20	24	29	11	16	23
SFPQ	44	46	44	44	42	32	25	22	31	32	24	31	34	40	39	30	35
	13	12	44	59	140	63	43	48	61	67	67	103	56	52	90	13	136
DDB1	56	48	43	36	34	31	18	33	16	25	19	19	26	29	20	8	27
PABPC1	43	36	42	18	33	15	15	16	1	15	20	14	8	23	21	5	10
PHB2	41	31	37	19	33	29	23	25	24	24	28	23	21	21	23	13	21
HSPA1A	35	41	34	25	35	28	24	28	28	22	27	28	31	33	17	12	22
RPS3	30	31	32	25	30	33	23	32	16	29	29	16	11	24	22	36	18
PKM	34	29	31	30	29	42	35	33	13	32	31	27	21	28	31	13	15
RPS5	41	37	29	19	21	22	14	7	2	11	22	5	0	6	28	24	12
DDOST	29	34	29	22	23	14	12	4	4	8	21	0	0	12	16	5	9
NOLC1	0	0	28	0	0	0	0	0	0	0	0	0	0	0	0	0	0
MYL6	29	21	25	11	17	8	10	7	8	6	9	10	9	12	8	5	7
RPSA	26	26	24	25	24	24	18	22	16	22	24	24	16	24	16	13	34
RPL13	27	30	24	23	27	12	18	7	15	13	13	18	20	18	16	22	4
NDUFS3	6	14	22	9	18	12	9	13	2	7	13	6	2	10	8	1	5
EMG1	21	17	19	12	15	9	8	9	0	6	10	6	2	15	9	3	9
RPS7	23	10	18	7	9	0	0	1	0	0	2	0	0	1	9	1	2
RPLP2	30	20	18	8	13	7	5	7	6	6	16	15	4	23	13	9	16
ERCC3	1	0	18	0	0	0	0	0	0	0	0	0	0	0	0	0	0
RUVBL1	13	18	17	6	16	4	7	4	1	0	10	1	2	2	2	0	2
RPL23	14	19	17	23	12	11	16	11	13	11	12	12	9	14	9	8	10
RFC4	13	8	17	3	10	5	2	0	2	5	2	6	4	5	4	0	3
COX7A2	15	15	17	15	15	7	9	7	0	8	11	10	0	4	10	5	7



Nucleolar and coiled-body  
phosphoprotein 1

Mr. N

VIM	369	314	314	167	303	70	127	88	84	92	105	85	71	91	145	90	127
RTRAF	105	103	94	97	84	66	71	55	13	63	78	60	32	83	76	28	70
SRP14	66	57	66	46	56	49	41	44	20	39	46	25	17	38	48	51	37
ACTB	65	57	64	51	43	20	38	24	27	28	44	22	13	28	37	25	39
PRPF19	50	32	61	44	39	38	16	20	5	14	27	16	15	40	25	12	58
NONO	59	71	61	52	70	29	28	16	15	42	35	12	22	19	29	52	48
PHB	66	48	60	30	52	42	33	30	17	31	47	27	17	40	50	25	39
RPS18	69	56	54	47	56	25	36	26	25	41	49	21	24	25	39	35	33
NUDT21	50	54	49	21	49	33	31	30	18	27	35	18	7	32	32	13	32
HSPA8	42	40	46	28	36	21	21	18	23	14	21	20	24	29	11	16	23
SFPQ	44	46	44	44	42	32	25	22	31	32	24	31	34	40	39	30	35
	13	12	44	59	140	63	43	48	61	67	67	103	56	52	90	13	136
DDB1	56	48	43	36	34	31	18	33	16	25	19	19	26	29	20	8	27
PABPC1	43	36	42	18	33	15	15	16	1	15	20	14	8	23	21	5	10
PHB2	41	31	37	19	33	29	23	25	24	24	28	23	21	21	23	13	21
HSPA1A	35	41	34	25	35	28	24	28	28	22	27	28	31	33	17	12	22
RPS3	30	31	32	25	30	33	23	32	16	29	29	16	11	24	22	36	18
PKM	34	29	31	30	29	42	35	33	13	32	31	27	21	28	31	13	15
RPS5	41	37	29	19	21	22	14	7	2	11	22	5	0	6	28	24	12
DDOST	29	34	29	22	23	14	12	4	4	8	21	0	0	12	16	5	9
NOLC1	0	0	28	0	0	0	0	0	0	0	0	0	0	0	0	0	0
MYL6	29	21	25	11	17	8	10	7	8	6	9	10	9	12	8	5	7
RPSA	26	26	24	25	24	24	18	22	16	22	24	24	16	24	16	13	34
RPL13	27	30	24	23	27	12	18	7	15	13	13	18	20	18	16	22	4
NDUFS3	6	14	22	9	18	12	9	13	2	7	13	6	2	10	8	1	5
EMG1	21	17	19	12	15	9	8	9	0	6	10	6	2	15	9	3	9
RPS7	23	10	18	7	9	0	0	1	0	0	2	0	0	1	9	1	2
RPLP2	30	20	18	8	13	7	5	7	6	6	16	15	4	23	13	9	16
ERCC3	1	0	18	0	0	0	0	0	0	0	0	0	0	0	0	0	0
RUVBL1	13	18	17	6	16	4	7	4	1	0	10	1	2	2	2	0	2
RPL23	14	19	17	23	12	11	16	11	13	11	12	12	9	14	9	8	10
RFC4	13	8	17	3	10	5	2	0	2	5	2	6	4	5	4	0	3
COX7A2	15	15	17	15	15	7	9	7	0	8	11	10	0	4	10	5	7





# Anti-NVL autoantibodies in systemic sclerosis

Vulsteke et al.  
Jan 2023

Preruna-Prieto et al.  
Sep 2023

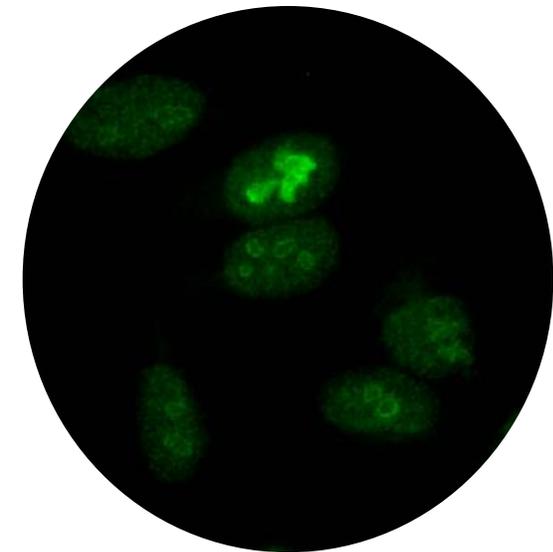
Matsuda et al.  
July 2023 (preprint)

	Vulsteke et al. Jan 2023	Preruna-Prieto et al. Sep 2023	Matsuda et al. July 2023 (preprint)	
<b>Number of patients</b>	<b>2</b>	<b>6</b>	<b>5</b>	<b>→ 13 cases</b>
Nucleolar HEp-2 IIFA pattern	2/2	6/6	5/5	→ Nucleolar pattern
Limited cutaneous	2/2	6/6	5/5	→ Limited cutaneous
Interstitial lung disease	0/2	2/6	0/5	→ Few ILD
Calcinosis	1/2	6/6	0/5	→ Unclear
Cancer or premalignancy	2/2	4/6	NA	→ <b>Potential association</b>

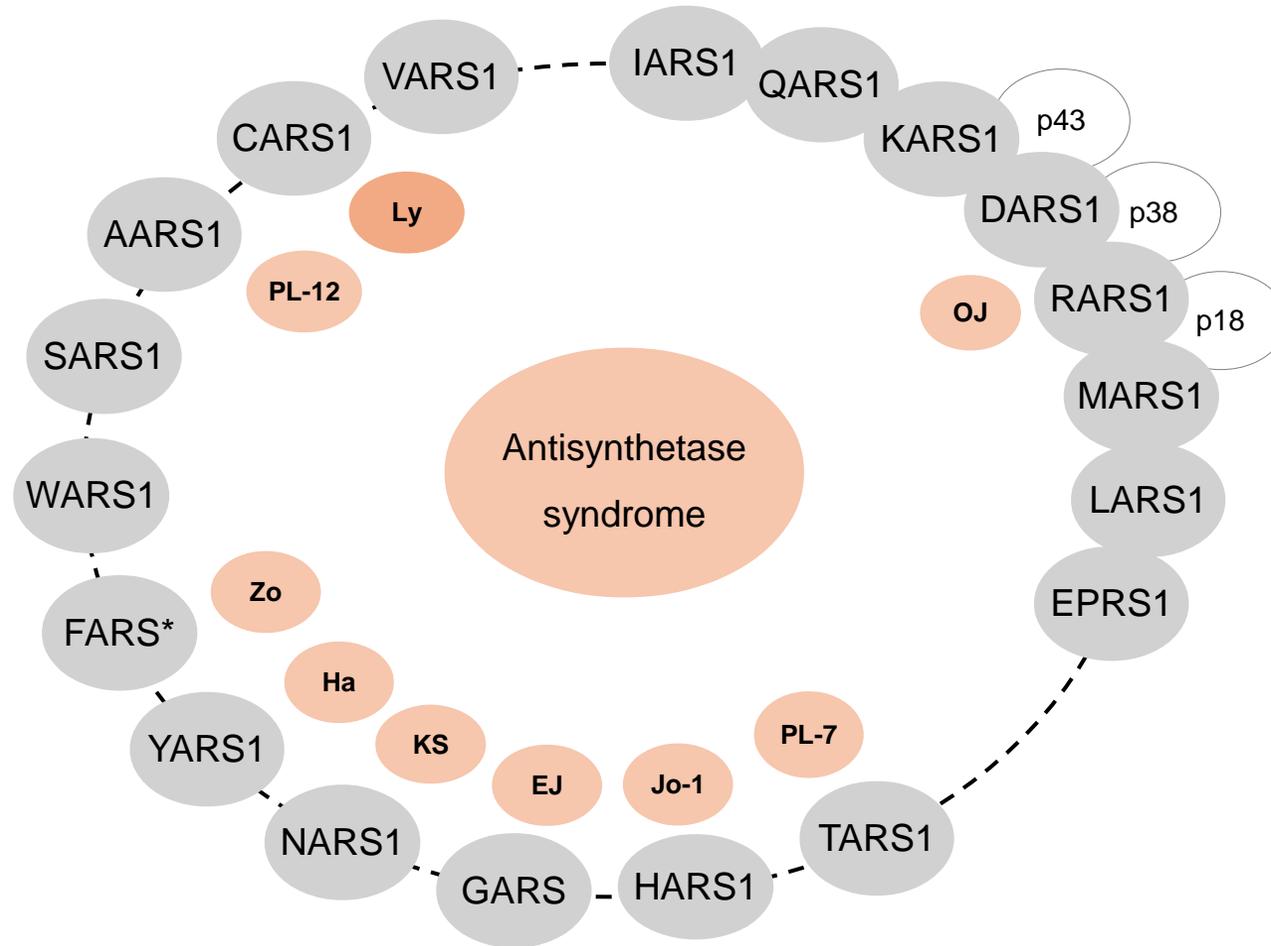
## Nuclear valosin-containing protein-like

*Telomerase assembly and function*

*Ribosome biogenesis*

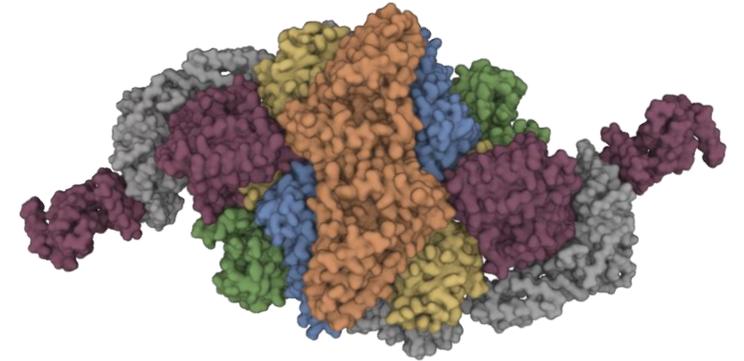


			Total spectral count per individual serum																				
			Possible ASyS, ASA-					Jo-1 (DIA)			PL-7 (DIA)		PL-12 (DIA)			EJ (DIA)			Healthy controls				
Identified protein	Abbr.	MW	U1	U2	U3	U4	U5	U6	U7	U8	U9	U10	U12	U13	U14	U15	U16	U17	U18	U19	U20	U21	
Cluster of Cysteinyl-tRNA synthetase	CARS1	85 kDa	51	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
Valine--tRNA ligase	VARS1	140 kDa	10	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
Bifunctional glutamate/proline--tRNA ligase	EPRS1	171 kDa	2	26	47	1	2	2	5	3	9	1	2	1	4	2	3	3	1	2	2	3	3
Glutamine--tRNA ligase	QARS1	88 kDa	0	18	28	0	2	1	1	1	1	1	0	1	2	0	0	1	1	0	1	0	0
Aspartate--tRNA ligase	DARS1	57 kDa	5	16	21	6	4	4	4	5	9	8	5	5	8	5	7	6	2	4	2	4	4
Lysine--tRNA ligase	KARS1	68 kDa	0	14	6	1	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0
Arginine--tRNA ligase	RARS1	75 kDa	3	13	43	3	2	2	3	1	2	4	3	2	5	2	1	2	1	2	0	0	0
Isoleucine--tRNA ligase	IARS1	145 kDa	1	8	19	1	1	1	1	1	3	0	1	0	5	1	0	0	1	2	0	0	0
Methionine--tRNA ligase	MARS1	101 kDa	0	2	6	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
Leucine--tRNA ligase	LARS1	134 kDa	0	0	67	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
Aminoacyl tRNA synthase complex-IMP1	AIMP1	34 kDa	2	10	11	1	2	1	0	2	4	0	1	1	1	0	1	1	0	0	2	0	0
Eukaryotic translation elongation factor 1 epsilon-1	EEF1E1	20 kDa	3	5	7	2	2	2	2	2	3	3	2	2	3	2	2	3	2	2	2	2	2
Aminoacyl tRNA synthase complex-IMP2	AIMP2	35 kDa	1	3	5	0	2	1	1	1	2	2	2	2	2	1	1	1	0	0	1	0	0
Phenylalanine--tRNA ligase alpha subunit	FARSA	58 kDa	2	2	3	28	2	2	2	2	2	2	1	1	1	2	2	2	2	2	2	2	2
Phenylalanine--tRNA ligase beta subunit	FARSB	66 kDa	0	0	0	26	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0
Asparagine--tRNA ligase	NARS1	63 kDa	0	0	0	0	22	0	0	4	1	0	0	0	1	0	0	0	0	0	0	0	0
Histidine--tRNA ligase	HARS1	57 kDa	0	2	0	0	0	32	21	21	0	0	0	0	0	0	0	0	0	0	0	0	0
Cluster of Threonine--tRNA ligase 1	TARS1	83 kDa	1	1	2	1	3	0	2	2	100	58	3	1	2	1	1	1	3	2	3	2	2
Alanine--tRNA ligase	AARS1	107 kDa	0	0	0	0	0	0	0	0	0	0	48	57	60	1	0	0	0	0	0	0	0
Glycine--tRNA ligase	GARS1	83 kDa	0	0	1	1	1	1	1	0	1	1	0	0	0	15	1	23	0	0	0	0	0
Acetyl-CoA carboxylase 1	ACACA	266 kDa	36	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
Cluster of ATPase family protein 2 homolog	SPATA5	98 kDa	0	0	0	0	11	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
Spermatogenesis-associated protein 5-like protein 1	SPATA5L1	81 kDa	0	0	0	0	11	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
Uncharacterized protein C1orf109	C1ORF109	23 kDa	0	0	0	0	7	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
RuvB-like 1	RUVBL1	50 kDa	1	5	3	1	3	5	1	2	3	3	2	1	4	4	72	3	0	2	1	4	4
RuvB-like 2	RUVBL2	51 kDa	1	2	2	3	2	3	3	2	3	1	2	3	3	2	71	3	2	3	1	3	3



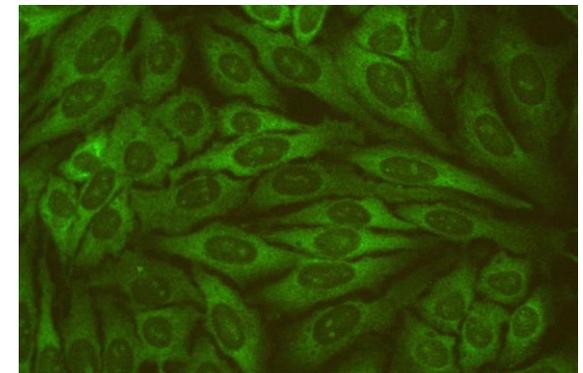
# eIF2B

		MW	#1	#2	#3	#4	#5	#6	H1	H2
eIF2B alpha	EIF2B1	34 kDa	5	4	7	225	42	103	0	0
eIF2B beta	EIF2B2	39 kDa	3	28	19	17	21	3	0	0
eIF2B gamma	EIF2B3	50 kDa	9	25	33	11	13	4	1	1
eIF2B delta	EIF2B4	60 kDa	18	41	38	16	9	11	0	0
eIF2B epsilon	EIF2B5	43 kDa	13	23	27	23	5	8	0	1

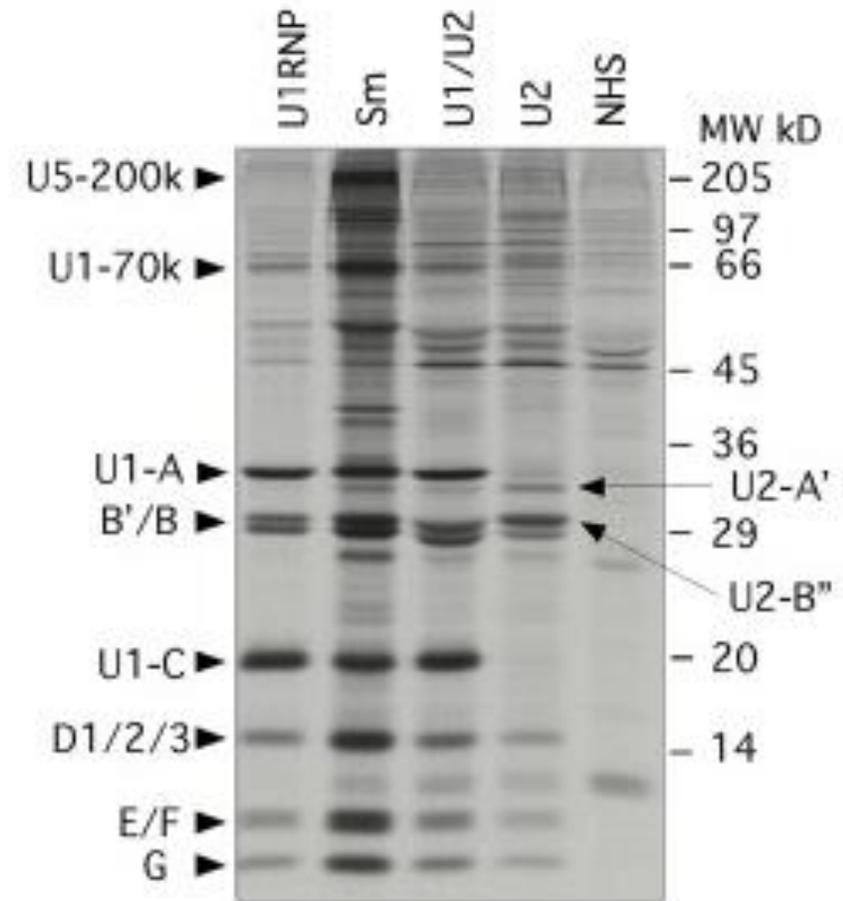
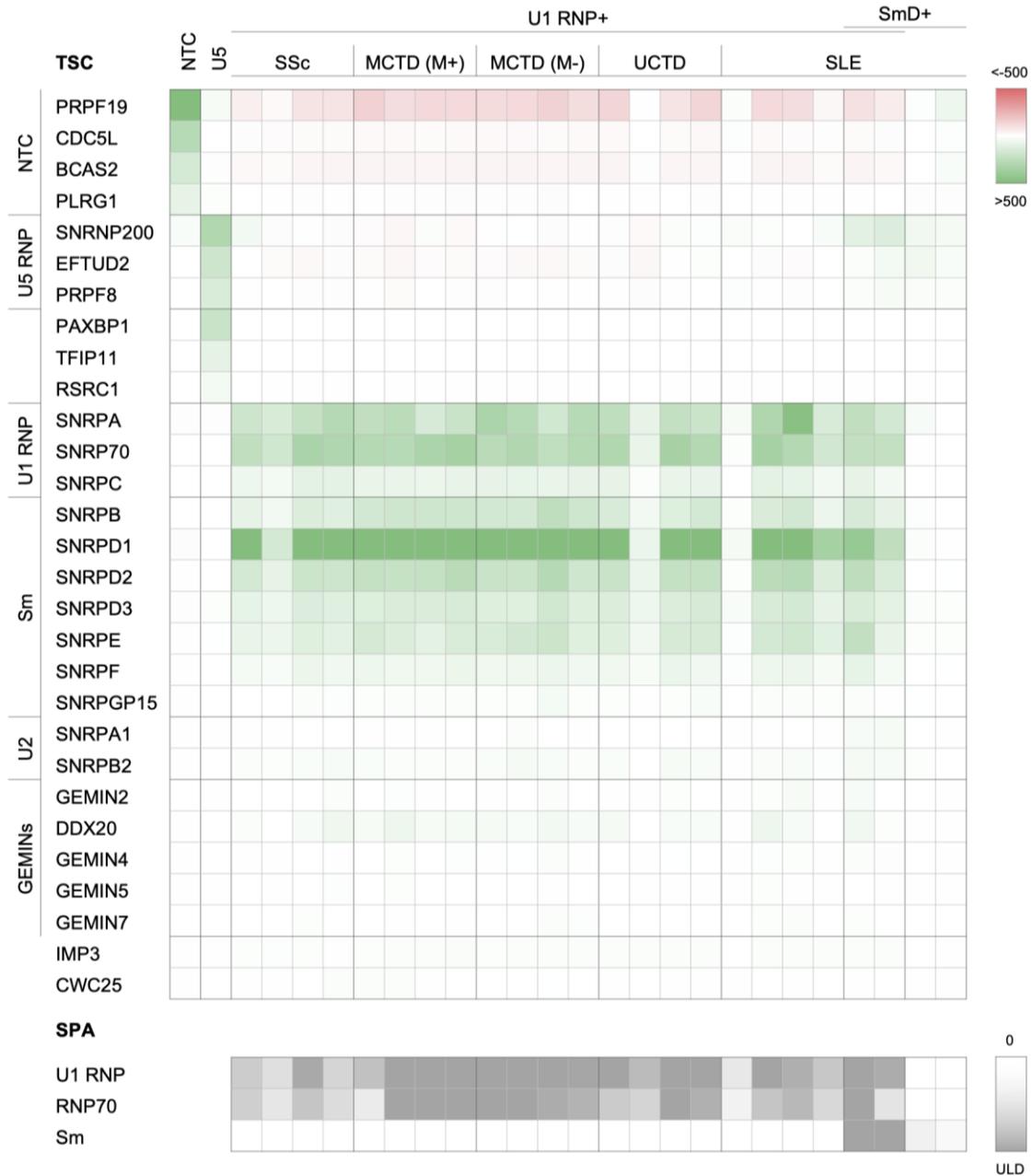


Diffuse cutaneous involvement  
 Interstitial lung disease  
 Severe cardiac involvement  
 Arthritis  
 Myositis

	■	■		■	
	■	■	■	■	■
	■			■	
■	■	■	■		
■	■			■	



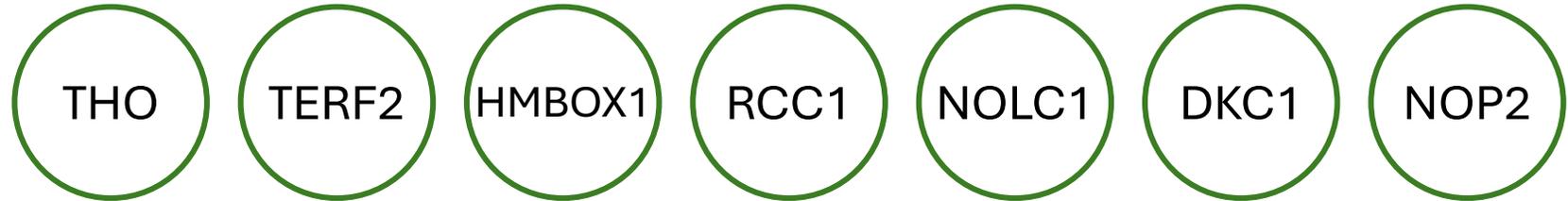
Kenner et al., Science 2019  
 Vulsteke et al., Rheumatology 2023



Vulsteke et al., J Autoimmun 2023  
Sato et al., Autoantibodies 2014

# The story of IP-MS thus far

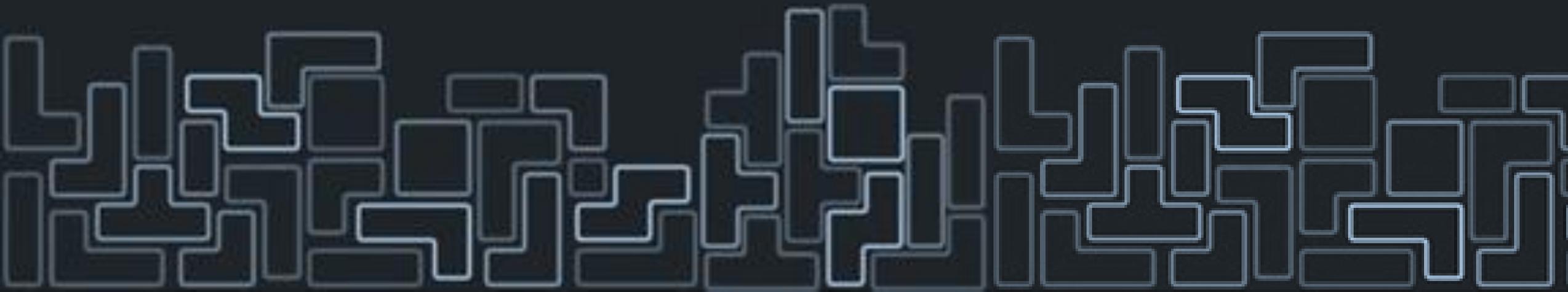
New

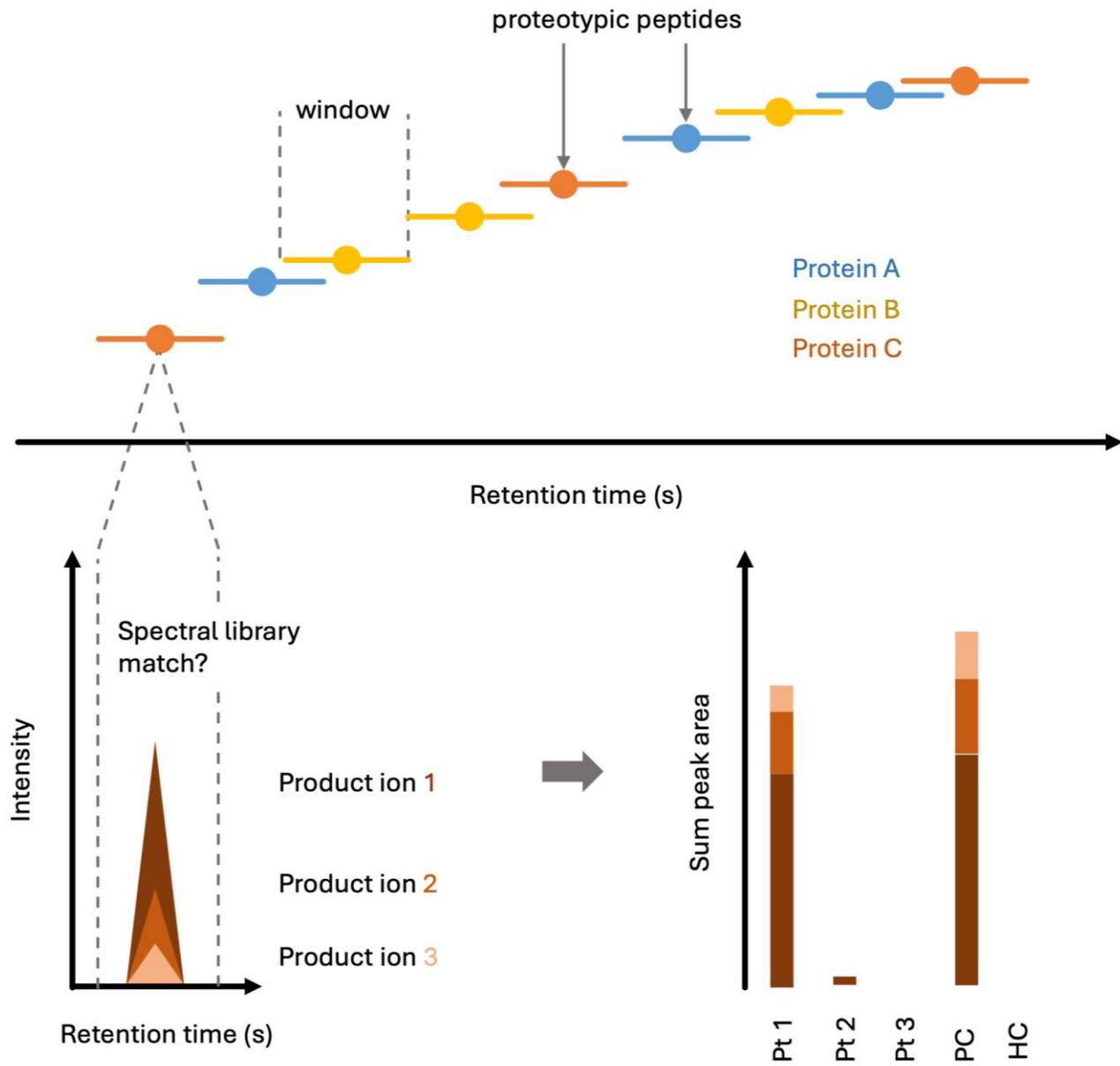


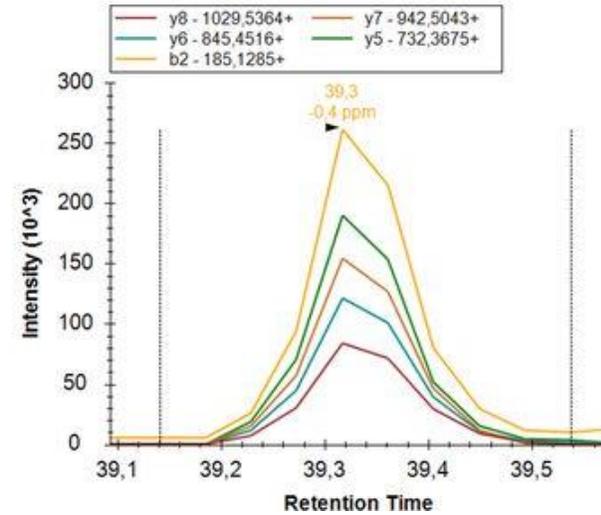
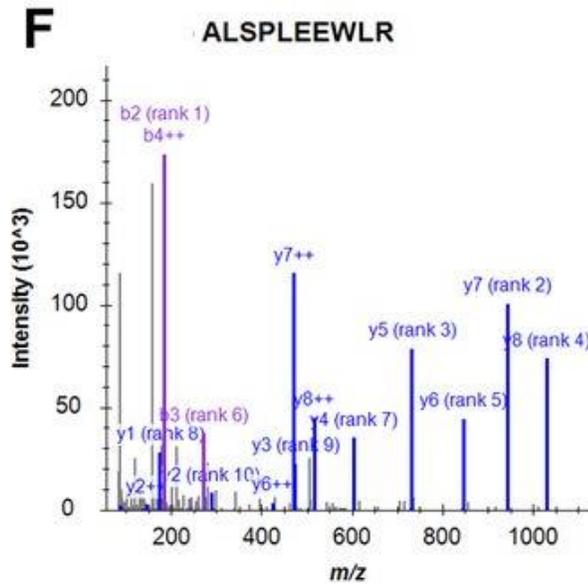
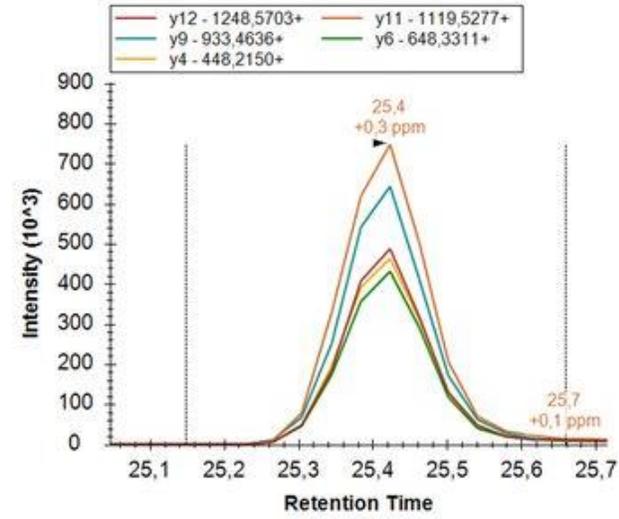
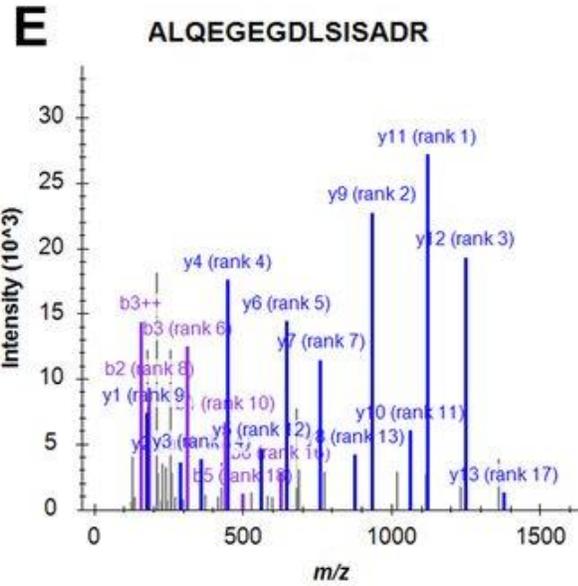
Rare

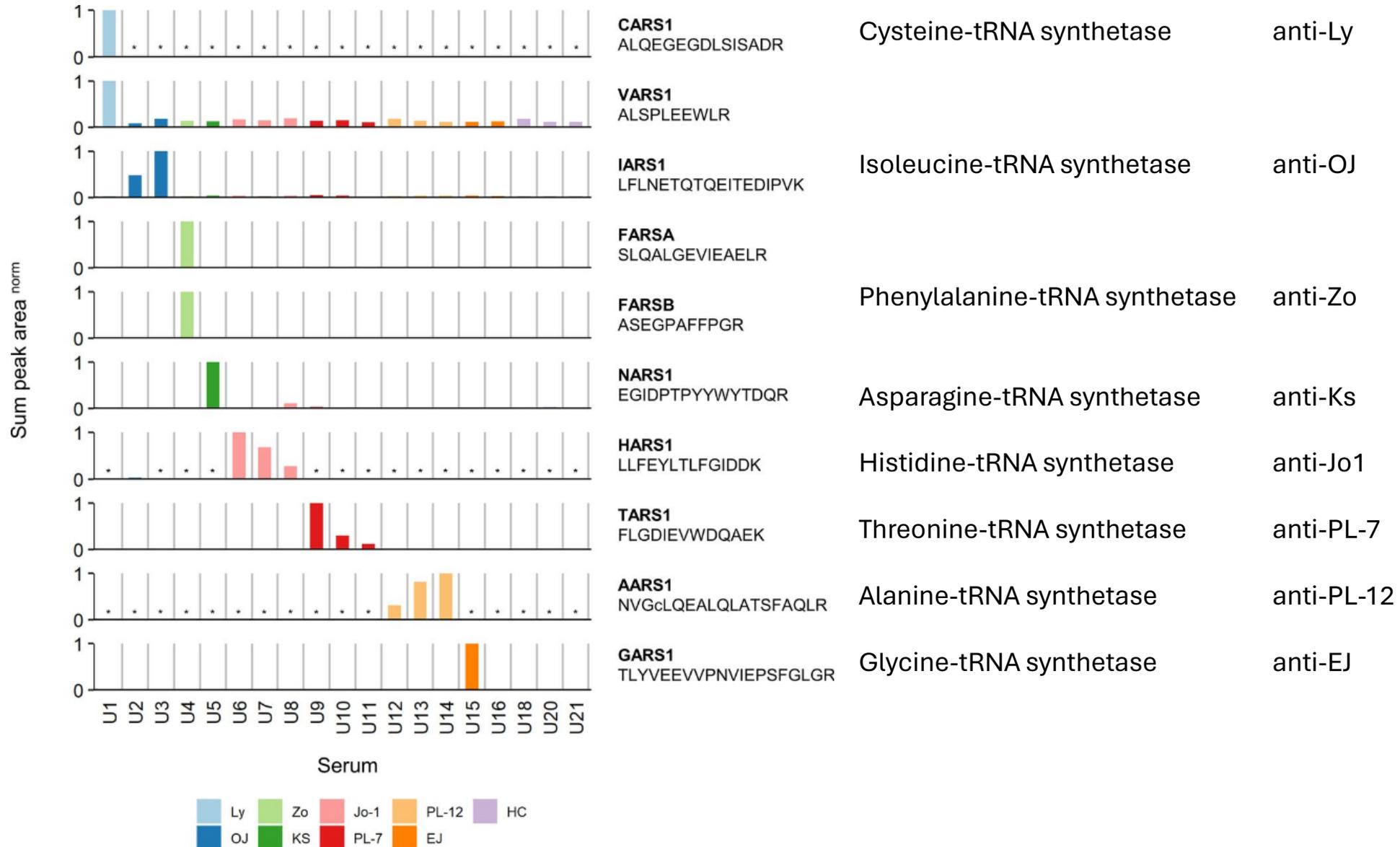


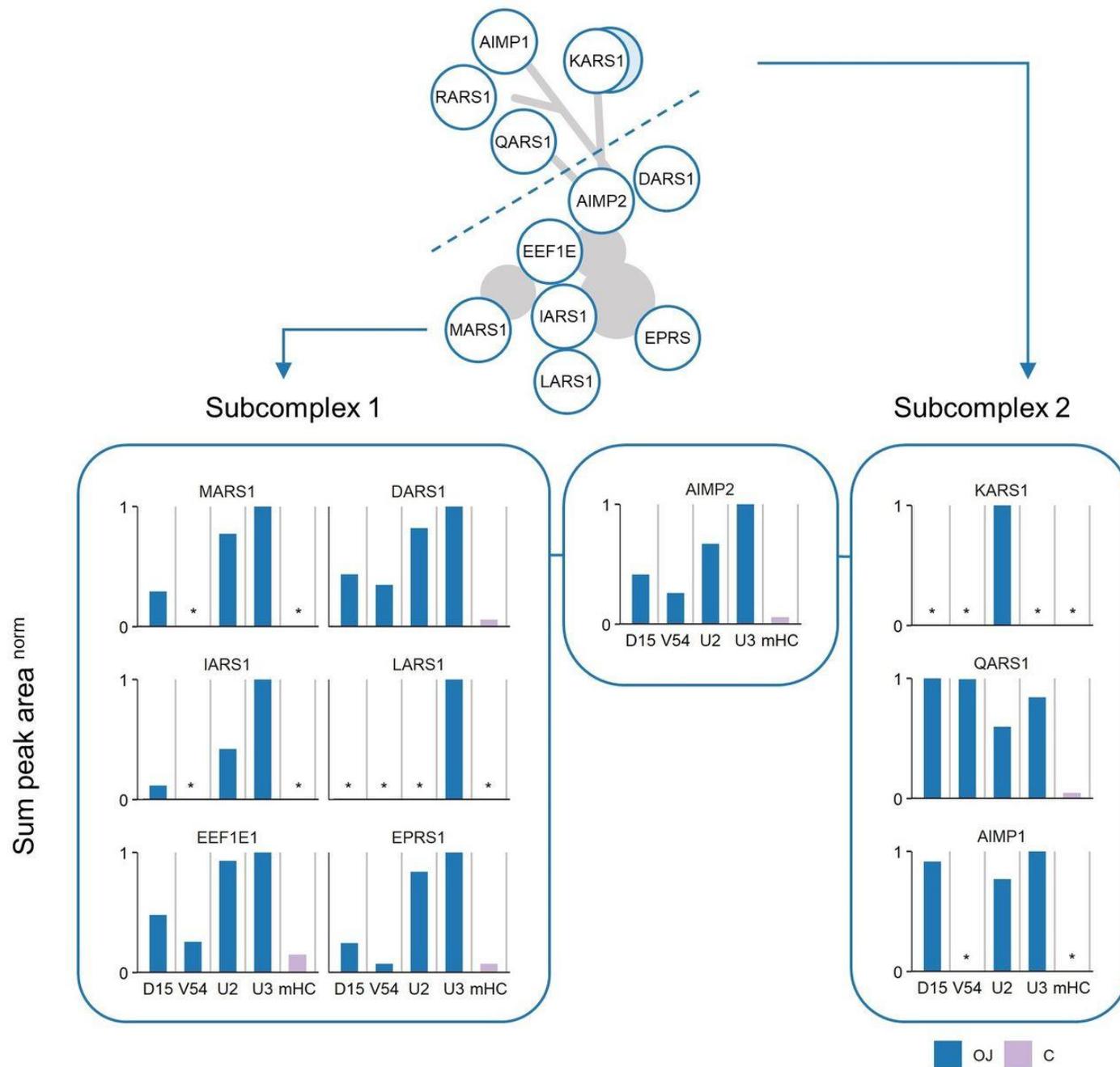
# Targeted IP-MS: one step further

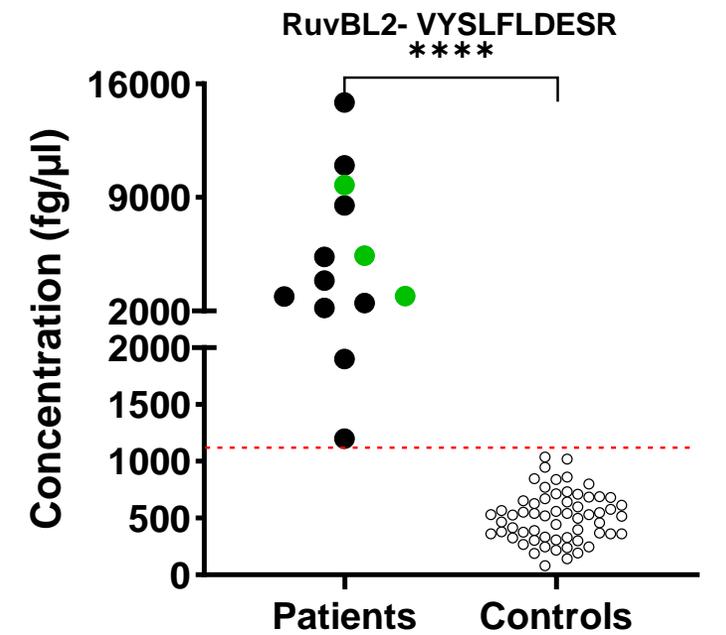
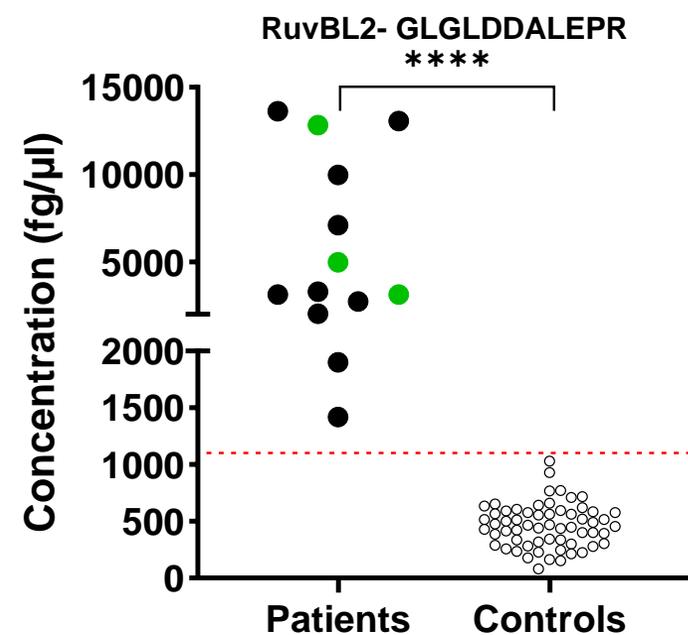
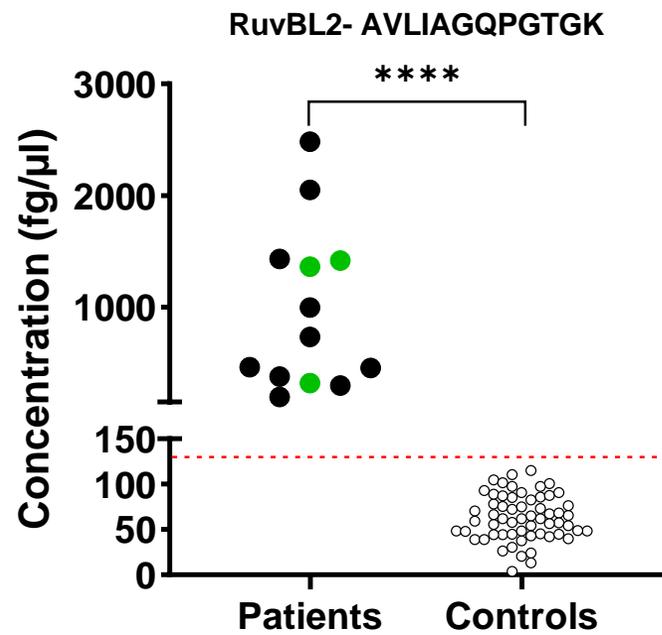
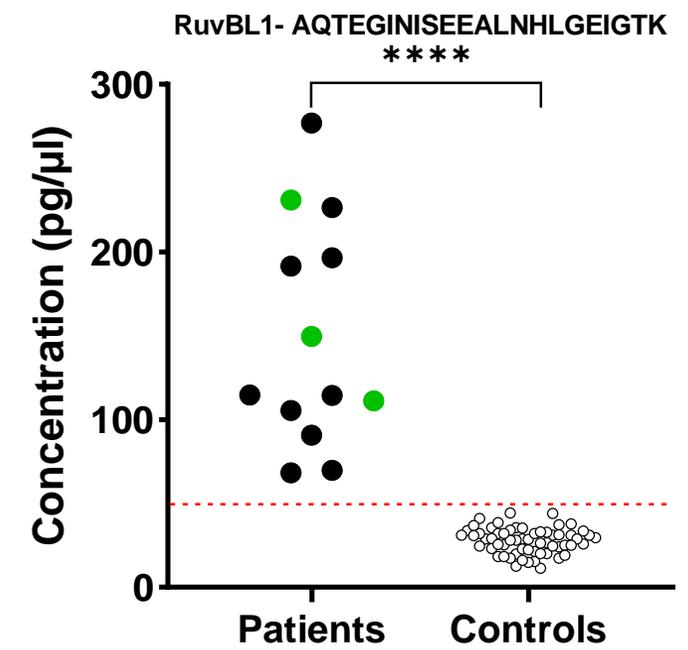
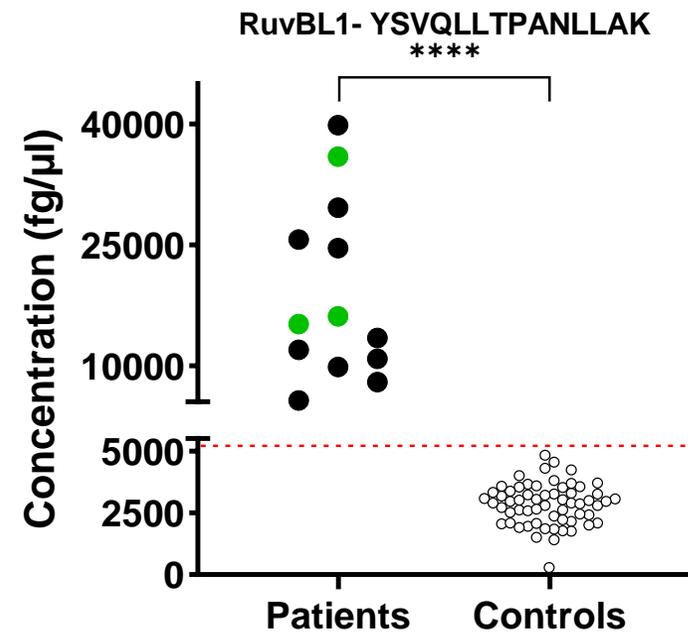
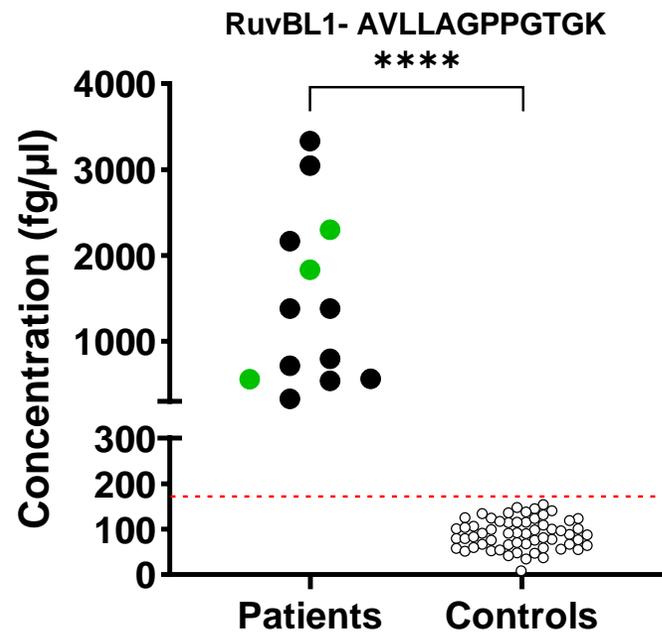












(n = 13 anti-RuvBL1/2 patients, n = 60 Disease Controls)



Nuclear HeLa extract 1



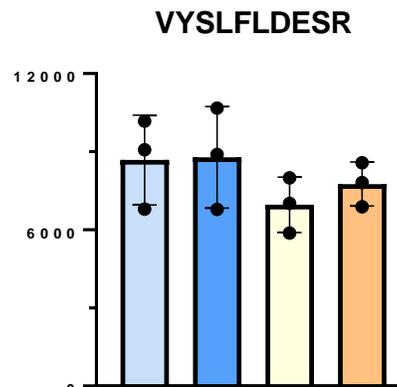
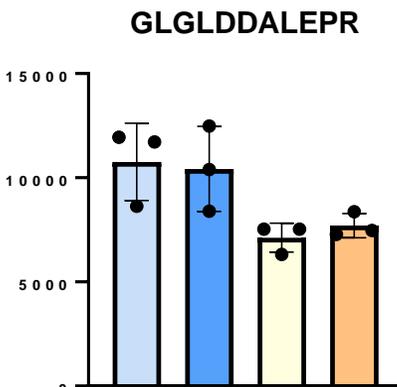
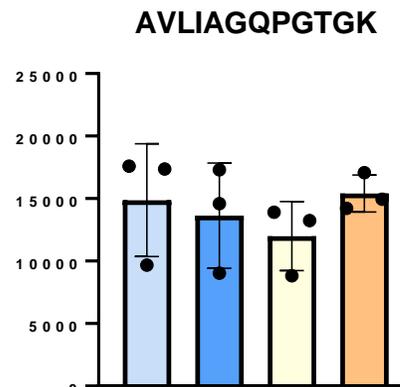
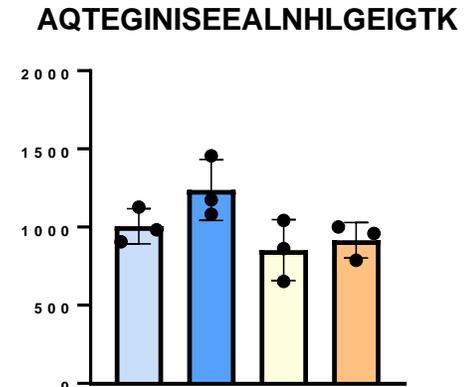
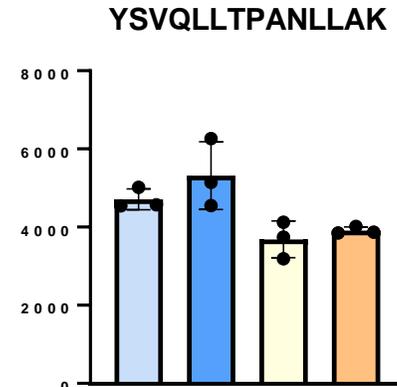
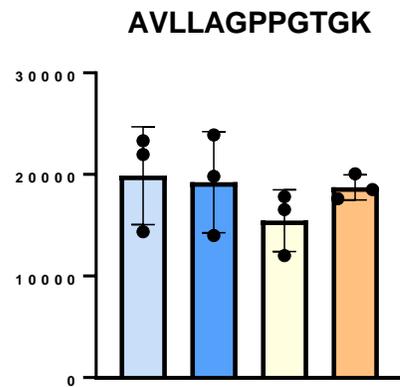
Cytoplasmic HeLa extract 1



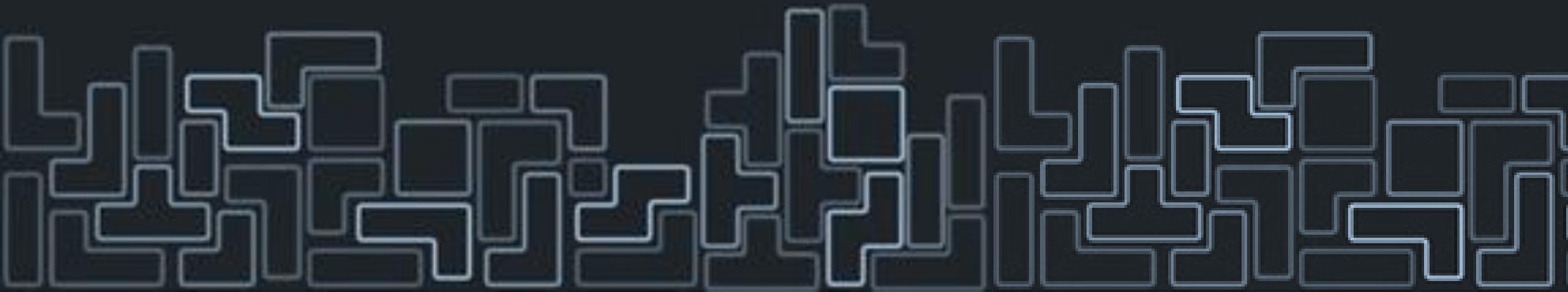
Nuclear HeLa extract 2

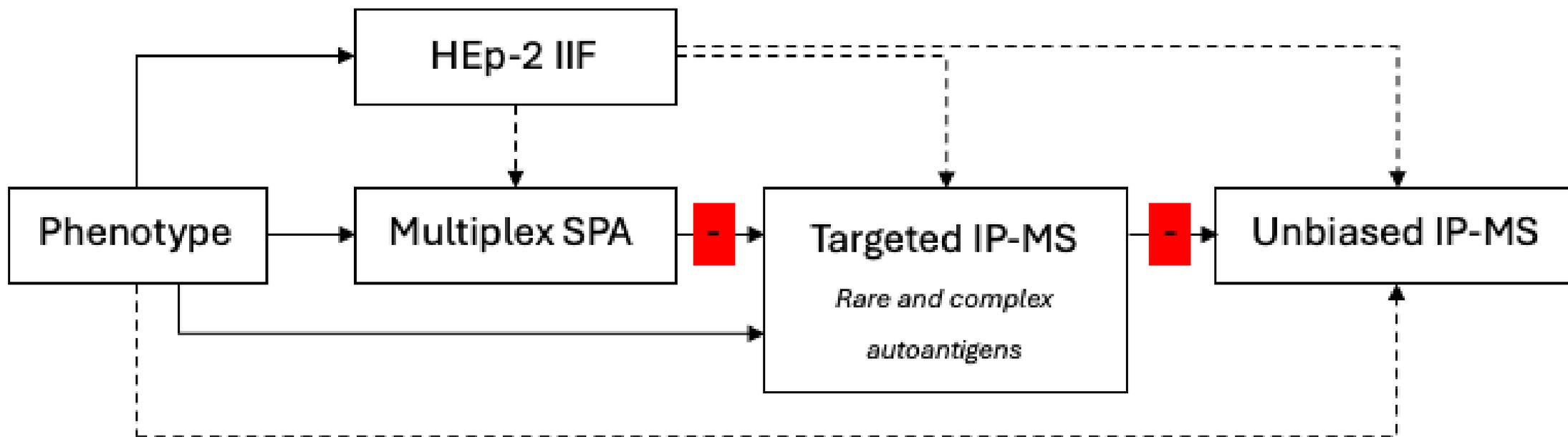


Cytoplasmic HeLa extract 2



**Is IP-MS ready for routine diagnostics?**







???

???

???

???

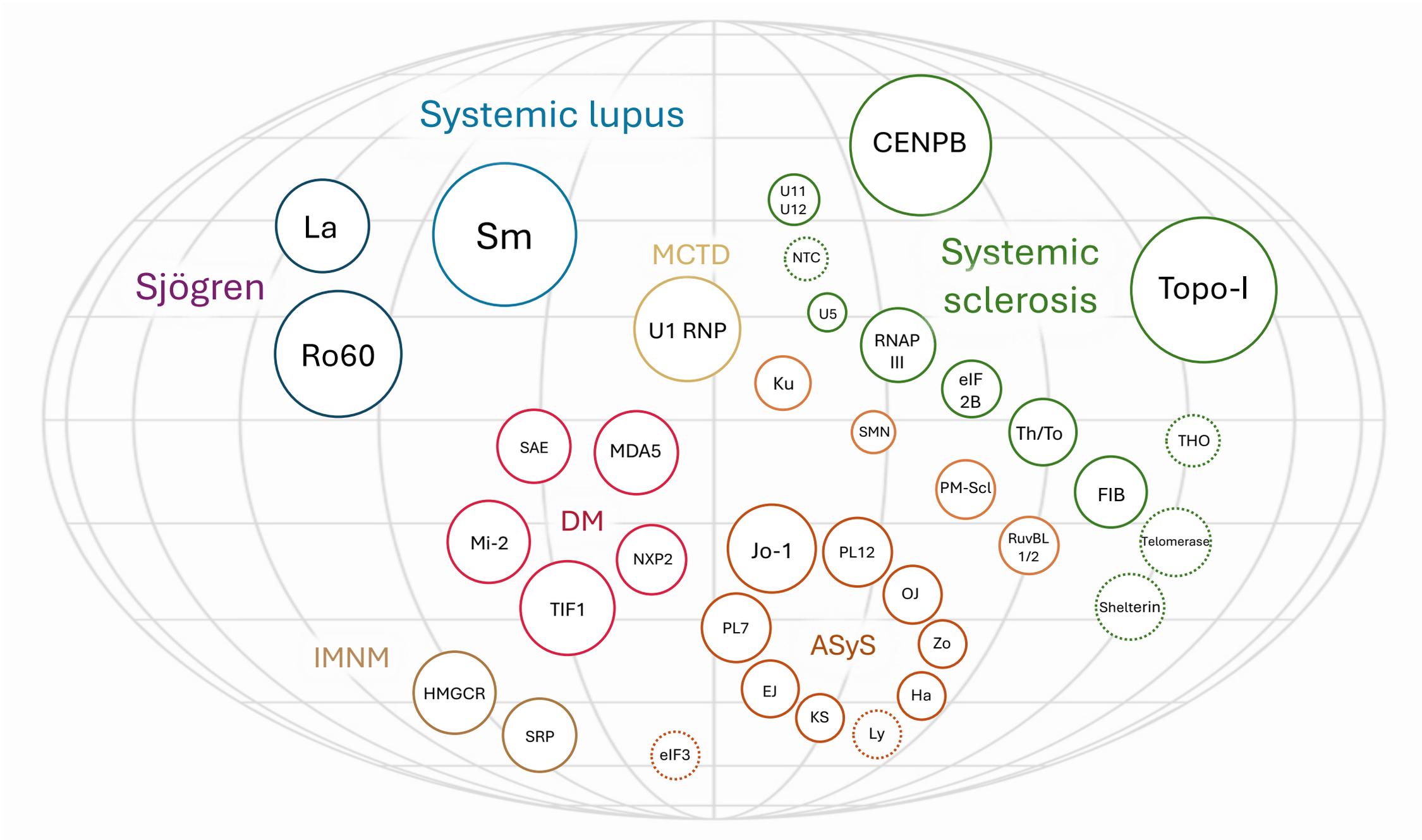
???

# BEL AIR



The **Belgian**

Autoantigen  
Identification  
Registry



Systemic lupus

Sjögren

La

Sm

Ro60

MCTD

U1 RNP

U11  
U12

NTC

U5

Ku

RNAP  
III

SMN

eIF  
2B

Th/To

Topo-I

Systemic  
sclerosis

THO

SAE

MDA5

Mi-2

DM

NXP2

TIF1

Jo-1

PL12

OJ

Zo

Ha

KS

EJ

PL7

ASyS

Ly

IMNM

HMGCR

SRP

eIF3

PM-Scl

FIB

Telomerase

Shelterin

RuvBL  
1/2



**KU LEUVEN**



**Fonds Joël Hurlet**

**Clinical and Diagnostic  
Immunology**

Xavier Bossuyt

Greet Wuyts

Doreen Dillaerts

Maaïke Cockx

Tom Dehaemer

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Rita Derua

Kusay Arat

Sebastien Carpentier

**Skeletal Biology Engineering  
and Research Center**

Ellen De Langhe

Rik Lories

Patrick Verschueren

**University Hospitals Leuven**

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Petra De Haes

Kristl Claeys

Jan Lenaerts

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Wim Wuyts

**Ghent University  
Hospital**

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